



# Turnitin Perpustakaan UKI

## NAVIGATING TUBERCULOSIS TREATMENT CHALLENGES OF GENETIC

-  Turnitin Dosen 29
-  Turnitin Dosen - Mei
-  Universitas Kristen Indonesia

---

### Document Details

**Submission ID**

trn:oid::1:3581703639

**Submission Date**

May 29, 2026, 1:35 PM GMT+7

**Download Date**

May 29, 2026, 2:05 PM GMT+7

**File Name**

NAVIGATING\_TUBERCULOSIS\_TREATMENT\_CHALLENGES\_OF\_GENETIC.pdf

**File Size**

250.7 KB

12 Pages

5,423 Words

35,218 Characters

# 13% Overall Similarity

The combined total of all matches, including overlapping sources, for each database.

## Filtered from the Report

- ▶ Bibliography

## Exclusions

- ▶ 2 Excluded Sources
- ▶ 8 Excluded Matches

## Match Groups

- 47 Not Cited or Quoted** 10%  
Matches with neither in-text citation nor quotation marks
- 12 Missing Quotations** 3%  
Matches that are still very similar to source material
- 0 Missing Citation** 0%  
Matches that have quotation marks, but no in-text citation
- 0 Cited and Quoted** 0%  
Matches with in-text citation present, but no quotation marks

## Top Sources

- 10% Internet sources
- 10% Publications
- 2% Submitted works (Student Papers)

## Integrity Flags

### 0 Integrity Flags for Review

No suspicious text manipulations found.

Our system's algorithms look deeply at a document for any inconsistencies that would set it apart from a normal submission. If we notice something strange, we flag it for you to review.

A Flag is not necessarily an indicator of a problem. However, we'd recommend you focus your attention there for further review.

### Match Groups

- **47 Not Cited or Quoted** 10%  
Matches with neither in-text citation nor quotation marks
- **12 Missing Quotations** 3%  
Matches that are still very similar to source material
- **0 Missing Citation** 0%  
Matches that have quotation marks, but no in-text citation
- **0 Cited and Quoted** 0%  
Matches with in-text citation present, but no quotation marks

### Top Sources

- 10% Internet sources
- 10% Publications
- 2% Submitted works (Student Papers)

### Top Sources

The sources with the highest number of matches within the submission. Overlapping sources will not be displayed.

|    |                |                                                                                        |     |
|----|----------------|----------------------------------------------------------------------------------------|-----|
| 1  | Publication    | Parissa Farnia, Ali Akbar Velayati, Jalaledin Ghanavi, Poopak Farnia. "Proteins in ... | <1% |
| 2  | Internet       | www.frontiersin.org                                                                    | <1% |
| 3  | Internet       | repositorio.upch.edu.pe                                                                | <1% |
| 4  | Publication    | Irene Oinike, Dewi Wijaya. "Immunology of Drug-Resistant Tuberculosis (DR-TB):...      | <1% |
| 5  | Internet       | www.mdpi.com                                                                           | <1% |
| 6  | Internet       | www.dovepress.com                                                                      | <1% |
| 7  | Internet       | mro.massey.ac.nz                                                                       | <1% |
| 8  | Internet       | public-pages-files-2025.frontiersin.org                                                | <1% |
| 9  | Student papers | University of KwaZulu-Natal                                                            | <1% |
| 10 | Publication    | Jen-Tsung Chen. "AI and Precision Medicine in Infectious Disease Management", ...      | <1% |

|    |             |                                                                                       |     |
|----|-------------|---------------------------------------------------------------------------------------|-----|
| 11 | Internet    | jurnal.globalhealthsciencegroup.com                                                   | <1% |
| 12 | Internet    | www.datainsightsmarket.com                                                            | <1% |
| 13 | Internet    | www.medicoverhospitals.in                                                             | <1% |
| 14 | Internet    | www.cdc.gov                                                                           | <1% |
| 15 | Publication | Rusana Rusana, Novita Anggraenni, Iwan Purnawan, Yuli Widyastuti, Risti Linta C...    | <1% |
| 16 | Publication | "Pathogenicity and Drug Resistance of Human Pathogens", Springer Science and ...      | <1% |
| 17 | Internet    | crbb-journal.com                                                                      | <1% |
| 18 | Publication | "Tuberculosis", Springer Science and Business Media LLC, 2023                         | <1% |
| 19 | Publication | Raj Kumar Mandal. "Nanobots and Pharmacological Interventions: A Future Pers...       | <1% |
| 20 | Internet    | bulletinofscience.kazatu.edu.kz                                                       | <1% |
| 21 | Internet    | www.eurpepsoc.com                                                                     | <1% |
| 22 | Publication | Hlanze, Hleliwe. "The Evolution of Drug Resistance in the Different Lineages of th... | <1% |
| 23 | Publication | Ndhlovu, Victor Sengelwayo. "Understanding Molecular Mechanisms of Mycobact...        | <1% |
| 24 | Publication | Tiana Carina Schwab, Lisa Perrig, Pauline Carlotta Göller, Freddy Fernando Guebe...   | <1% |

|    |             |                                                                                        |     |
|----|-------------|----------------------------------------------------------------------------------------|-----|
| 25 | Internet    | pmc.ncbi.nlm.nih.gov                                                                   | <1% |
| 26 | Internet    | www.ijebmr.com                                                                         | <1% |
| 27 | Internet    | www.preprints.org                                                                      | <1% |
| 28 | Internet    | www.science.org                                                                        | <1% |
| 29 | Internet    | www2.mdpi.com                                                                          | <1% |
| 30 | Publication | Dilpreet Singh, Vrinda Krishna, Nitya Kumari, Anoushka Banerjee, Prithviraj Kapo...    | <1% |
| 31 | Publication | Peter D O Davies, Stephen B Gordon, Geraint Davies. "Clinical Tuberculosis", CRC ...   | <1% |
| 32 | Internet    | edoc.unibas.ch                                                                         | <1% |
| 33 | Internet    | www.science.gov                                                                        | <1% |
| 34 | Publication | Yang Zhang, Ruiying Wu, Mingrui Sun, Xiaotian Li, Ren Fang, Jiayin Xing, Zhaoli Li,... | <1% |
| 35 | Publication | Raden Muhammad Indra, Irsan Saleh, Irawan Mangunatmadja, Yulia Iriani, Zen H...        | <1% |



## NAVIGATING TUBERCULOSIS TREATMENT CHALLENGES OF GENETIC PERSPECTIVES AND DNA-BASED DETECTION: A LITERATURE REVIEW

Rouly Pola Pasaribu<sup>1,7</sup>, Zen Hafy<sup>2\*</sup>, Ariesti Karmila<sup>3,7</sup>, Nur Riviaty<sup>4,7</sup>, Zen Ahmad<sup>1,7</sup>, Fadhyl Zuhry Lubis<sup>5,7</sup>, Welly Salutondok<sup>6</sup>

<sup>1</sup>Division of Pulmonology and Critical Medicine, Department of Internal Medicine, Faculty of Medicine, Universitas Sriwijaya, Jl. Suka Jaya No.319, Sukajaya, Sukarami, Palembang, Sumatera Selatan 30151 Indonesia

<sup>2</sup>Biomedical Department, Faculty of Medicine, Universitas Sriwijaya, Jl. Suka Jaya No.319, Sukajaya, Sukarami, Palembang, Sumatera Selatan 30151 Indonesia

<sup>3</sup>Division of Infectious and Tropical Disease, Department of Child Health, Faculty of Medicine, Universitas Sriwijaya, Jl. Suka Jaya No.319, Sukajaya, Sukarami, Palembang, Sumatera Selatan 30151 Indonesia

<sup>4</sup>Division of Geriatrics, Department of Internal Medicine, Faculty of Medicine, Universitas Sriwijaya, Jl. Suka Jaya No.319, Sukajaya, Sukarami, Palembang, Sumatera Selatan 30151 Indonesia

<sup>5</sup>Internal Medicine, Department of Internal Medicine; Faculty of Medicine, Universitas Sriwijaya, Jl. Suka Jaya No.319, Sukajaya, Sukarami, Palembang, Sumatera Selatan 30151 Indonesia

<sup>6</sup>Internal Medicine, Department of Internal Medicine; Faculty of Medicine, Universitas Kristen Indonesia, Jl. Mayor Jendral Sutoyo No.2, Cawang, Kramat jati, Jakarta Timur, Jakarta 13630, Indonesia

<sup>7</sup>Dr Mohammad Hoesin General Hospital, Jl. Jend. Sudirman Km. 3,5, Sekip Jaya, Kemuning, Palembang, Sumatera Selatan 30126, Indonesia

\*[zenhafy@gmail.com](mailto:zenhafy@gmail.com)

### ABSTRACT

Tuberculosis remains a major global health problem, particularly because of the increasing burden of drug-resistant tuberculosis (DR-TB), including MDR-TB and XDR-TB. Resistance in *Mycobacterium tuberculosis* is driven mainly by spontaneous genetic mutations, adaptive evolution, tolerance mechanisms, and host microenvironmental factors, which complicate both treatment and diagnosis. This review aimed to examine recent advances in the genetic mechanisms underlying antibiotic resistance in *M. tuberculosis* and to evaluate the progress of DNA-based detection technologies for tuberculosis diagnosis and management. This literature review analyzed 49 scientific articles published between 2012 and 2026 to examine the molecular and genetic basis of drug resistance in *M. tuberculosis*. The study focused on identifying target gene mutations, efflux pump mechanisms, compensatory evolution, and heteroresistance. Furthermore, it evaluated the clinical application of advanced DNA-based diagnostic tools, including GeneXpert, line probe assays, targeted next-generation sequencing (tNGS), whole-genome sequencing (WGS), droplet digital PCR, and CRISPR-based detection. Information was qualitatively synthesized to compare the efficacy and limitations of these modern diagnostic strategies in global tuberculosis management. Drug resistance was primarily associated with de novo mutations in genes such as *rpoB*, *katG*, *inhA*, *pncA*, *gyrA/gyrB*, and *rrs/eis*, as well as compensatory mutations that preserve bacterial fitness. In addition, non-genetic tolerance, dormancy, and efflux pump overexpression contributed to bacterial survival. DNA-based technologies enabled faster and more comprehensive resistance detection than conventional methods, although limitations remained in sensitivity, mutation coverage, cost, and infrastructure requirements. Genetic mutations, evolutionary adaptation, and tolerance mechanisms form the core basis of drug resistance in *M. tuberculosis*. Integrating DNA-based diagnostics into clinical practice is essential to improve early resistance detection, support personalised treatment, and reduce the spread of drug-resistant tuberculosis.

Keywords: drug resistance; DNA detection; efflux pump; genetic mutation; mycobacterium tuberculosis; whole-genome sequencing

### How to cite (in APA style)

Pasaribu, R. P., Hafy, Z., Karmila, A., Riviaty, N., Ahmad, Z., Lubis, F. Z., & Salutondok, W. (2026). Navigating Tuberculosis Treatment Challenges of Genetic Perspectives and DNA-Based Detection: A Literature Review. *Indonesian Journal of Global Health Research*, 8(4), 143–154. <https://doi.org/10.37287/ijghr.v8i4.1926>.

## INTRODUCTION

Tuberculosis (TB) remains one of the most significant infectious diseases worldwide, causing substantial morbidity and mortality despite the availability of effective treatment regimens (Nimmo, C et al., 2022). The disease is caused by *Mycobacterium tuberculosis*, a pathogen that continues to pose a major challenge to global public health due to its ability to persist, spread, and develop resistance to antitubercular drugs (Anes, E., et al., 2023). The growing emergence of drug-resistant tuberculosis (DR-TB), including multidrug-resistant TB (MDR-TB) and extensively drug-resistant TB (XDR-TB), has made TB control increasingly difficult (Pal, R., Bisht, M. K., & Mukhopadhyay, S., 2022). Resistance to first-line and second-line drugs limits treatment options, prolongs therapy, increases costs, and worsens patient outcomes. In many settings, delayed diagnosis and inappropriate empirical treatment further contribute to ongoing transmission and the expansion of resistant strains (Borah, P. et al., 2021).

At the molecular level, resistance in *M. tuberculosis* is driven mainly by spontaneous genetic mutations rather than horizontal gene transfer. Mutations in genes such as *rpoB*, *katG*, *inhA*, *pncA*, *gyrA*, and *gyrB* alter drug targets or activation pathways, reducing the effectiveness of major antitubercular agents (Pshennikova, E. S., & Voronina, A. S., 2022). In addition, non-genetic mechanisms such as efflux pump overexpression, dormancy, and phenotypic tolerance also support bacterial survival under antibiotic pressure (Canalda-Baltrons, A et al., 2025). Recent advances in DNA-based diagnostic technologies have improved the ability to detect TB and identify resistance more rapidly and accurately. (MacGregor-Fairlie et al., 2020) Methods such as GeneXpert, line probe assays, targeted next-generation sequencing, whole-genome sequencing, droplet digital PCR, and CRISPR-based diagnostics are transforming TB diagnosis and surveillance. These tools offer important opportunities for earlier detection, more precise treatment selection, and better control of resistant TB (Chauhan, M et al., 2024).

This review discusses the genetic basis of drug resistance in *M. tuberculosis* and highlights the role of modern DNA-based detection methods in improving TB management. By understanding these molecular mechanisms and diagnostic advances, more effective strategies can be developed to address the ongoing challenge of drug-resistant tuberculosis (Datta, D et al., 2024). This review discusses the genetic basis of drug resistance in *M. tuberculosis* and highlights the role of modern DNA-based detection methods in improving TB management. The objective of this study is to examine the recent advances in genetic mechanisms underlying antibiotic resistance specifically mutations in target genes and compensatory evolution and to evaluate the progress of DNA-based detection technologies, such as NGS and CRISPR, in enhancing diagnostic accuracy and personalized treatment for drug-resistant tuberculosis. By understanding these molecular mechanisms and diagnostic advances, more effective strategies can be developed to address the ongoing challenge of drug-resistant tuberculosis.

## METHOD

This study employed a literature review design to examine the genetic basis of antibiotic resistance in *Mycobacterium tuberculosis* and the evolution of contemporary DNA-based diagnostic approaches. Data collection involved a systematic search of reputable scientific databases, including PubMed, Scopus, and Google Scholar. The selected literature comprised empirical research and review articles published between 2012 and 2026, ensuring the integration of the most recent advancements in genomic technology. (de Araujo, L et al., 2023). From an initial identification of 65 relevant publications, 49 articles were selected as primary sources based on their thematic alignment and methodological rigor. The review focused specifically on mutations within resistance-associated target genes such as *rpoB*, *katG*, *inhA*, *pncA*, *gyrA*, *gyrB*, *rrs*, and *eis* as well as broader mechanisms including efflux pump activity, compensatory evolution, dormancy, and heteroresistance. Additionally, the analysis evaluated the clinical application of advanced molecular diagnostic tools, including GeneXpert, line probe assays, targeted next-generation sequencing

(tNGS), whole-genome sequencing (WGS), droplet digital PCR, and CRISPR-based detection methods. (Dheda, K et al., 2024). All gathered information was qualitatively synthesized using a thematic approach to compare the strengths, limitations, and clinical relevance of each resistance mechanism and diagnostic efficacy. The findings were organized systematically to provide a comprehensive overview of early detection strategies and the future management of drug-resistant tuberculosis. (Dousa, K. M et al., 2020).

## RESULT

### Biological and Genetic Foundations of *M. tuberculosis*

The biological resilience of *M. tuberculosis* relies heavily on its unique and highly complex cell envelope, which creates a formidable barrier against both the host immune system and antibiotic penetration. This envelope features a core composed of a mycolic acid-peptidoglycan-arabinogalactan (mAGP) complex.<sup>11</sup> The long, highly hydrophobic mycolic acid chains (C60-C90) are particularly critical for the bacterium's virulence and persistence, forming a thick, waxy protective layer that drastically limits cellular permeability. Above this core lies a peripheral lipid layer containing vital components like phthiocerol dimycocerosates (PDIM) and trehalose dimycolate (TDM), topped by an outer capsule of alpha-glucan and arabinomannan. This structural dynamic naturally confers a high degree of intrinsic resistance to the pathogen, explaining the necessity for long-term and complex therapeutic regimens to successfully eradicate the bacteria.

Beyond its physical defenses, *M. tuberculosis* utilizes a sophisticated array of virulence factors and proteins to manipulate the host's cellular environment, particularly within macrophages. A primary weapon is the ESX-1 Type VII secretion system, which facilitates the release of effector proteins such as the Early Secretory Antigenic Target 6 (ESAT-6) and Culture Filtrate Protein 10 (CFP-10).<sup>14,15</sup> ESAT-6 functions as a porin that can lyse the phagosomal membrane, allowing the bacteria to escape degradation in the phagolysosome and enter the nutrient-rich cytoplasm of the host cell. Meanwhile, CFP-10 stabilizes ESAT-6 to ensure efficient secretion while also modulating host immune responses. Additionally, the bacterium dedicates nearly ten percent of its coding capacity to the PE (Proline-Glutamic acid) and PPE (Proline-Proline-Glutamic acid) protein families. These proteins are localized on the cell surface or secreted to subvert host immunity and dictate the fate of the infected macrophage, pushing it toward either protective apoptosis or tissue-damaging necroptosis depending on the stage of infection.

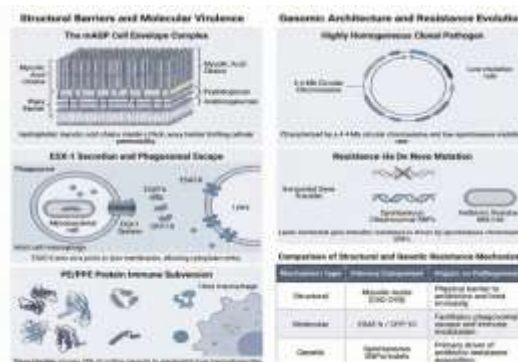


Figure 1. Mycobacterium Tuberculosis Pathogenesis and Resilience (Gengenbacher, M., & Kaufmann, S. H., 2012).

### Molecular Mechanisms of Antibiotic Resistance

The molecular mechanisms underlying antibiotic resistance in *M. tuberculosis* are exceptionally complex and multifactorial, driven primarily by specific spontaneous genetic mutations that alter drug targets or interfere with drug activation pathways, as well as physiological adaptations to the host environment. Unlike many bacteria that rely on horizontal gene transfer, *M. tuberculosis* acquires hereditary resistance primarily through de novo chromosomal mutations such as single nucleotide polymorphisms (SNPs), insertions, and deletions which become fixed in the population

under the selective pressure of antibiotic therapy. The clinical consequences of these mutations depend heavily on the specific gene involved, the resulting increase in the minimum inhibitory concentration (MIC), and the overarching genomic context of the bacterial strain (Gygli, S. M., et al 2017).

Table 1.  
 Molecular Mechanisms and Diagnostic Profiles of Antibiotic Resistance in TB

| Antibiotic Class            | Target Gene/Locus | Resistance Mechanism                                                                         | Diagnostic Methodology                                       | Clinical Classification | Fitness Cost & Compensatory Evolution (Inferred)                                                         |
|-----------------------------|-------------------|----------------------------------------------------------------------------------------------|--------------------------------------------------------------|-------------------------|----------------------------------------------------------------------------------------------------------|
| Rifamycins                  | rpoB              | Target modification; RNA polymerase beta-subunit prevents drug binding                       | WGS, targeted sequencing (Deeplex-MycTB), Xpert MTB/RIF, LPA | MDR-TB, XDR-TB          | Low fitness cost for S450L; compensatory mutations in rpoA or rpoC often restore fitness.                |
| Isonicotinic acids          | katG              | Loss of pro-drug activation; prevents activation by catalase-peroxidase enzyme               | WGS, Deeplex-MycTB, Xpert MTB/XDR, LPA                       | MDR-TB, XDR-TB, HR-TB   | Minimal fitness cost; allows enzyme to maintain essential defensive functions against host ROS.          |
| Diarylquinolines/Phenazines | mmpR5 (Rv0678)    | Efflux pump overexpression; loss of repressor function leads to MmpL5/MmpS5 pump activity    | WGS, targeted sequencing (Deeplex-MycTB)                     | XDR-TB                  | Moderate fitness cost; cross-resistance between Bedaquiline and Clofazimine often observed.              |
| Isonicotinic acids          | inhA (promoter)   | Target modification/overexpression; reduces drug affinity and increases enzyme concentration | WGS, Deeplex-MycTB, Xpert MTB/XDR, LPA                       | MDR-TB, HR-TB           | Generally lower fitness cost compared to katG deletions, leading to higher transmissibility.             |
| Fluoroquinolones            | gyrA / gyrB       | Target modification; prevents drug binding to DNA gyrase                                     | WGS, Deeplex-MycTB, Xpert MTB/XDR, LPA                       | Pre-XDR, XDR-TB         | Some mutations in gyrA carry low fitness costs; compensatory mutations can occur in gyrB.                |
| Aminoglycosides             | rrs, eis          | Target modification (rrs) or enzymatic inactivation (eis acetyltransferase)                  | WGS, LPA, Xpert MTB/XDR                                      | XDR-TB                  | rrs mutations can significantly reduce translation efficiency; eis mutations often allow higher fitness. |
| Pyrazinamide                | pncA              | Loss of pro-drug activation; loss of pyrazinamidase (PZase) enzyme function                  | WGS, Deeplex-MycTB                                           | MDR-TB                  | High variability in fitness cost depending on the specific mutation in the pncA gene.                    |
| Nitroimidazoles             | ddn               | Loss of pro-drug activation; results in non-functional proteins for Delamanid activation     | WGS                                                          | MDR/XDR-TB              | Variable; resistance emerges under selective pressure from prior drug exposure.                          |

**Evolutionary Adaptation and Genetic Diversity**

Unlike many other bacterial species that rapidly acquire new survival traits through horizontal gene transfer, Mycobacterium tuberculosis rarely utilizes this mechanism to adapt to its environment.<sup>26</sup> Integrating foreign genetic material carries a high risk of disrupting vital enzymatic functions, and such material is generally eliminated via purifying selection. Consequently, the evolutionary adaptation and acquisition of antibiotic resistance in M. tuberculosis rely almost entirely on the accumulation of spontaneous de novo chromosomal mutations that provide a selective advantage under the heavy pharmacological pressure of antibiotic therapy. While mutations conferring resistance typically impose a biological “fitness cost” that reduces the bacterium’s overall viability and replication speed, M. tuberculosis has evolved to elegantly counteract this burden through compensatory mutations. For example, the highly prevalent rpoB S450L mutation, which causes resistance to rifampicin, imposes minimal fitness loss when it is accompanied by secondary

3

compensatory mutations in the *rpoA* or *rpoC* genes. These secondary mutations stabilize the RNA polymerase complex and fully restore the bacterium’s cellular fitness, allowing multidrug-resistant strains to survive, maintain high transmissibility, and spread efficiently across human populations without a biological disadvantage.

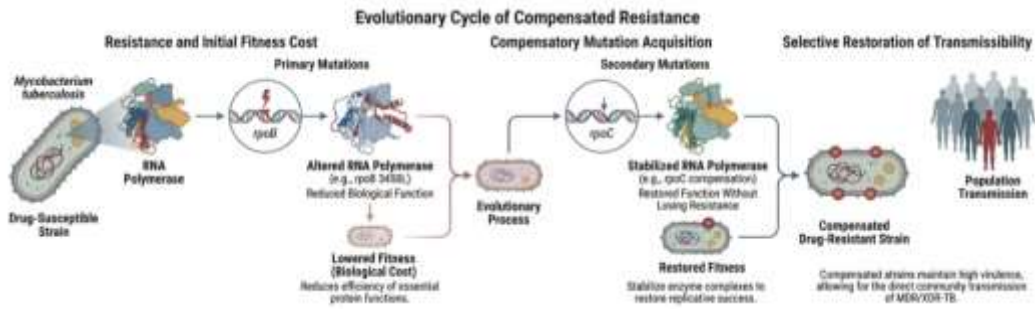


Figure 2. Evolutionary Dynamics of Fitness Compensation in Drug-Resistant Mycobacterium tuberculosis (Jose Vadakunnel, M et al., 2025)

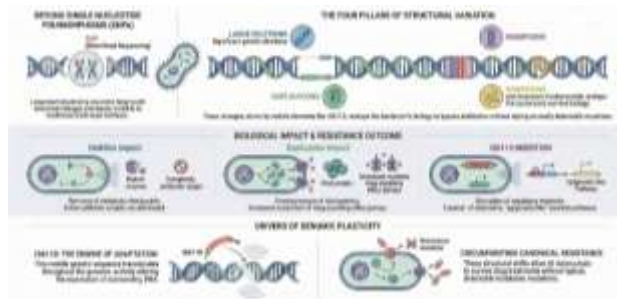


Figure 3. The Impact of IS6110-Driven Genomic Rearrangements on Survival Pathways and Resistance in Mycobacterium tuberculosis (Katoch, V. M., 2020)

**Advancements in DNA-Based Detection Technologies**

Table 2.

Comparison of current & emerging molecular diagnostic technologies for Mycobacterium tuberculosis

| Kategori                        | Nama Teknologi          | Keunggulan Utama                                                             | Keterbatasan Utama                                                       | Target Penanda                                                                    |
|---------------------------------|-------------------------|------------------------------------------------------------------------------|--------------------------------------------------------------------------|-----------------------------------------------------------------------------------|
| Cepat & Praktis (Point-of-Care) | Xpert MTB/RIF & XDR     | Sangat cepat (<2 jam); mendeteksi bakteri sekaligus resistensi obat utama.   | Biaya cartridge tinggi; hanya mendeteksi mutasi yang sudah diketahui.    | <i>rpoB</i> , <i>katG</i> , <i>inhA</i> , <i>gyrA/B</i> , <i>rrs</i> , <i>eis</i> |
|                                 | CRISPR-Cas12a           | Sangat sensitif; hasil visual tanpa alat rumit; waktu <1,5 jam.              | Masih tahap riset; risiko efek <i>off-target</i> .                       | IS6110                                                                            |
| Sekuensing (Komprehensif)       | Targeted NGS            | Profil resistensi lengkap untuk 15 jenis obat; identifikasi ratusan spesies. | Tidak dapat mendeteksi mutasi baru di luar panel target.                 | 15 Gen Resistensi                                                                 |
|                                 | Whole Genome Sequencing | Standar emas; memetakan seluruh genom bakteri secara utuh.                   | Waktu lama (~72 jam); biaya tinggi; butuh analisis bioinformatika rumit. | Seluruh Genom                                                                     |
|                                 | Oxford Nanopore         | Alat portabel; mampu membaca struktur varian DNA yang kompleks.              | Angka kesalahan pembacaan lebih tinggi dibanding Illumina.               | Seluruh Genom                                                                     |
| Lainnya                         | Line Probe Assay (LPA)  | Efektif untuk skrining lini pertama & kedua secara manual.                   | Padat karya; interpretasi hasil bersifat subjektif.                      | <i>rpoB</i> , <i>katG</i> , <i>inhA</i> , <i>gyrA/B</i> , <i>rrs</i> , <i>eis</i> |

## Genotype-Phenotype Discordance and Heteroresistance

Table 3.

Genotype-Phenotype Discordance and Heteroresistance

| Aspect                         | Findings                                                                                                                                                                                                             | Clinical Implication                                                                                               |
|--------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------|
| Genotype-phenotype discordance | DNA-based tests may detect resistance-associated mutations that do not always produce clear phenotypic resistance. Some isolates may appear susceptible in culture-based testing despite being genetically resistant | This may lead to underestimation of resistance and inappropriate treatment selection.                              |
| Low-level resistance           | Certain mutations produce low-level resistance that may fall below the detection threshold of conventional phenotypic assays.                                                                                        | Patients may receive ineffective standard therapy, increasing the risk of treatment failure pmc.ncbi.nlm.nih.      |
| Heteroresistance               | Resistant and susceptible subpopulations can coexist in the same patient, with resistant fractions reported as low as 1% to 10% in some studies sciencedirect.                                                       | Minority resistant variants may be missed by routine testing, especially when bacterial load is low sciencedirect. |
| Conventional methods           | Phenotypic drug susceptibility testing may fail to identify low-frequency resistant subpopulations.                                                                                                                  | Resistance may be recognised late, after resistance has expanded under treatment pressure                          |
| Molecular methods              | Targeted NGS and ddPCR can detect mixed populations and low-frequency mutations more sensitively than conventional assays.                                                                                           | These methods improve early detection of emerging resistance and support more precise treatment decisions.         |
| Overall impact                 | Discordance and heteroresistance complicate interpretation of TB drug-resistance results.                                                                                                                            | Integrated molecular and phenotypic assessment is needed for accurate diagnosis and regimen selection.             |

## Clinical Challenges and Future Directions

Table 4.

Clinical Challenges and Future Directions

| Aspect                                     | Description                                                                                              | Implication                                                                                    |
|--------------------------------------------|----------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------|
| Resistance to newer drugs                  | Resistance is also emerging against bedaquiline, clofazimine, delamanid, and pretomanid.                 | Treatment options become more limited and complex.                                             |
| Diagnostic difficulty in paucibacillary TB | Extrapulmonary TB, paediatric TB, and TB in HIV-coinfected patients often have very low bacterial loads. | Conventional tests may miss the infection or its resistance pattern.                           |
| Delay in appropriate therapy               | Slow or incomplete diagnosis can lead to empirical treatment.                                            | This increases the risk of treatment failure and ongoing transmission.                         |
| Need for molecular integration             | Rapid molecular triage and sequencing-based profiling should be integrated into routine care.            | Faster identification of resistance supports more accurate treatment selection.                |
| Tiered diagnostic strategy                 | Use rapid point-of-care tests first, followed by advanced sequencing in reference laboratories.          | This approach is practical and cost-effective, especially in low- and middle-income countries. |
| Drug optimisation                          | Molecular results can help avoid unnecessary use of toxic second-line drugs.                             | Reduces adverse effects and improves resource allocation.                                      |
| Future drug discovery                      | Genetic engineering and functional genomics can validate essential bacterial targets.                    | Accelerates development of new antitubercular drugs.                                           |
| Genomic surveillance                       | Ongoing monitoring of resistant strains and new mutations is needed.                                     | Supports personalised treatment and public health control.                                     |

## DISCUSSION

### Biological and Genetic Foundations of *M. tuberculosis*

Mycobacterium tuberculosis has a thick, lipid-rich cell wall that makes it highly resistant to host immune responses and the penetration of antitubercular drugs. Genetically, the bacterium survives and adapts mainly through spontaneous mutations and the expression of virulence factors that support infection, survival, and persistence within the human body.

### Molecular Mechanisms of Antibiotic Resistance

Resistance to primary first-line drugs, specifically rifampicin and isoniazid, constitutes the

2  
19

33

fundamental barrier to successful tuberculosis treatment and defines the multidrug-resistant (MDR-TB) phenotype. Rifampicin functions by binding to the RNA polymerase beta-subunit to arrest mRNA synthesis, a process disrupted almost exclusively by mutations within the *rpoB* gene's 81-base pair region, predominantly the S450L substitution. Conversely, isoniazid resistance involves a more intricate genetic profile; as a prodrug, it requires activation by the KatG enzyme. The dominant *katG* S315T mutation bypasses this activation while uniquely preserving the enzyme's ability to defend the pathogen against host-generated oxidative stress, ensuring bacterial survival without compromising its physiological fitness. The therapeutic landscape further complicates as resistance emerges against second-line agents and novel compounds, such as fluoroquinolones, bedaquiline, and clofazimine. Fluoroquinolone resistance, a hallmark of extensively drug-resistant tuberculosis (XDR-TB), arises from mutations in the *gyrA* and *gyrB* genes that prevent drug binding to DNA gyrase. Beyond these specific genetic modifications, *M. tuberculosis* employs non-specific biological defenses, including drug-inactivating enzymes and the overexpression of the MmpL5/MmpS5 efflux pump system. Crucially, through the DosR regulon, the bacteria can transition into a non-replicating dormant state within host granulomas. This physiological persistence renders drugs targeting active metabolism ineffective and serves as an evolutionary bridge, allowing sub-populations to survive long enough to acquire the permanent genetic mutations that dictate absolute resistance.

### Evolutionary Adaptation and Genetic Diversity

3

The resistance of *M. tuberculosis* is not merely a byproduct of individual genetic mutations but a result of sophisticated evolutionary adaptation. The pathogen exhibits significant genetic diversity, allowing sub-populations to survive diverse environmental pressures within the host. Central to this adaptation is the role of "fitness-neutral" mutations, such as the *rpoB* S450L substitution, which grants high-level resistance without compromising the bacterium's reproductive success. This stability is often reinforced by compensatory mutations in the *rpoA* or *rpoC* genes, which mitigate the physiological burden of resistance and ensure the long-term survival of multidrug-resistant strains.

Furthermore, the transition between genetic resistance and non-genetic tolerance represents a crucial evolutionary strategy. Through regulatory networks like the DosR regulon, the bacteria can enter a state of metabolic dormancy. This physiological persistence allows the pathogen to withstand prolonged antibiotic exposure in a non-replicating state, effectively serving as an evolutionary bridge. By surviving initial pharmacological assaults through these adaptive mechanisms, *M. tuberculosis* gains the necessary window to acquire permanent, high-level genetic mutations, thereby driving the continuous evolution of drug resistance.

### Advancements in DNA-Based Detection Technologies

5  
18  
5

The rapid evolution of DNA-based diagnostic tools has fundamentally shifted the paradigm of tuberculosis management from slow, culture-based methods to rapid molecular precision. Current advancements, particularly in Nucleic Acid Amplification Tests (NAATs) like Xpert MTB/RIF Ultra, allow for the simultaneous detection of *M. tuberculosis* and critical resistance markers within hours. The integration of Loop-mediated Isothermal Amplification (LAMP) further enhances diagnostic accessibility in resource-limited settings by eliminating the need for complex thermal cyclers, thereby bridging the gap between high-tech laboratory settings and point-of-care clinical practice. Furthermore, the emergence of Whole Genome Sequencing (WGS) and Targeted Next-Generation Sequencing (tNGS) offers an unprecedented depth of genetic insight. Unlike traditional assays that target specific loci, sequencing technologies provide a comprehensive profile of the bacterial genome, enabling the detection of rare or compensatory mutations that contribute to low-level resistance. This shift toward high-throughput genomic analysis not only improves individual treatment outcomes through personalized medicine but also enhances global surveillance of resistant strains. By identifying emerging resistance patterns in real-time, these DNA-based

technologies serve as a critical defense against the further escalation of XDR-TB and other highly resistant phenotypes.

### Genotype-Phenotype Discordance and Heteroresistance

16 A significant challenge in modern tuberculosis diagnostics is the occurrence of genotype-phenotype discordance, where molecular results do not align with traditional drug susceptibility testing (DST). This discrepancy often arises from "silent" or synonymous mutations that molecular assays may flag as resistance markers despite the bacteria remaining phenotypically sensitive. Conversely, discordance occurs when clinical resistance is present, yet the specific genetic locus targeted by the assay remains wild-type. This suggests the involvement of alternative resistance mechanisms, such as mutations in unmapped regulatory regions or the hyperactivation of non-specific efflux pumps, which underscores the inherent limitations of targeted molecular diagnostics compared to the comprehensive nature of phenotypic DST. The complexity of resistance is further intensified by heteroresistance, a phenomenon characterized by the co-existence of drug-susceptible and drug-resistant *M. tuberculosis* subpopulations within the same patient. Heteroresistance represents an early evolutionary stage of resistance, often invisible to conventional diagnostic methods if the resistant fraction falls below the assay's detection threshold. This intra-host diversity poses a severe clinical risk; under the selective pressure of antibiotic treatment, the resistant subpopulation can rapidly expand, leading to treatment failure and the eventual stabilization of a fully resistant genotype. Addressing these diagnostic gaps requires the integration of high-sensitivity sequencing technologies capable of detecting low-frequency variants to ensure more accurate and personalized therapeutic interventions

### Genotype-Phenotype Discordance and Heteroresistance

24 A significant challenge in modern tuberculosis diagnostics is genotype-phenotype discordance, where molecular results fail to align with traditional drug susceptibility testing (DST). This discrepancy manifests either as "false resistance" due to non-clinical synonymous mutations or, more critically, as "false sensitivity" where clinical resistance exists without mutations in targeted genetic regions. Such discordance suggests the involvement of alternative mechanisms, such as distal regulatory mutations or the hyperactivation of non-specific efflux pumps, which underscores the inherent limitations of current targeted molecular assays. The diagnostic landscape is further complicated by heteroresistance, the simultaneous presence of drug-susceptible and drug-resistant subpopulations within a single host. This intra-host diversity often remains undetected by standard DST if the resistant fraction is below the 1% threshold, leading to "stealth" resistance and treatment failure under antibiotic pressure. Addressing these gaps requires a shift toward high-sensitivity technologies like Targeted Next-Generation Sequencing (tNGS) and Deep Sequencing, which can detect low-frequency variants. Recognizing these nuances is a clinical imperative to prevent the amplification of resistance and improve outcomes for MDR-TB and XDR-TB patients.

### Clinical Challenges and Future Directions

16 The clinical management of drug-resistant tuberculosis (DR-TB) faces persistent hurdles despite significant technological breakthroughs. One of the primary clinical challenges is the complexity of interpreting molecular data in the presence of rare or novel mutations. As diagnostic reliance shifts toward genetic detection, clinicians often encounter "variants of unknown significance" (VUS)—mutations whose actual impact on drug efficacy remains uncharacterized. This uncertainty can lead to suboptimal dosing or the premature abandonment of effective drugs. Furthermore, the global disparity in laboratory infrastructure remains a critical barrier; while high-income regions utilize comprehensive genomic sequencing, many high-burden countries still rely on basic smear microscopy or limited molecular assays, leading to delayed diagnoses and continued community transmission. Another pressing challenge is the biological reality of persister cells and intracellular sequestration. Even when genetic testing suggests susceptibility, the physiological state of *M. tuberculosis* within the protective environment of a granuloma can shield it from pharmacological

action. Current therapeutic regimens often fail to account for this metabolic dormancy, necessitating prolonged treatment durations that contribute to high default rates and the subsequent amplification of resistance.

## CONCLUSION

Tuberculosis remains a profound global health crisis, heavily exacerbated by the rapid spread of multidrug-resistant and extensively drug-resistant strains. The remarkable resilience of *Mycobacterium tuberculosis* stems from its severe genetic plasticity, utilizing spontaneous chromosomal mutations, compensatory evolution to restore cellular fitness, and large-scale structural variants (such as gene deletions and duplications) to survive antibiotic pressure and evade traditional detection methods. To overcome these complex survival mechanisms, clinical management must shift toward advanced DNA-based diagnostic technologies. High-throughput platforms like Whole-Genome Sequencing (WGS), portable Nanopore devices, and ultra-sensitive CRISPR-Cas assays allow clinicians to rapidly map comprehensive resistance profiles and track localized transmission clusters in real-time. Simultaneously, cutting-edge genetic engineering tools such as the ORBIT recombineering system and multiplexed CRISPR interference (CRISPRi) are revolutionizing functional genomics by drastically accelerating the discovery and validation of novel therapeutic targets. By combining these advanced molecular innovations with emerging host-directed therapies that target host-pathogen protein interactions, and integrating them into modernized global health security frameworks, the international community can move decisively closer to the total eradication of tuberculosis.

## REFERENCES

- Anes, E., Pires, D., Mandal, M., & Azevedo-Pereira, J. M. (2023). ESAT-6 a major virulence factor of *Mycobacterium tuberculosis*. *Biomolecules*, 13(6), 968.
- Borah, P., Deb, P. K., Venugopala, K. N., Al-Shar'i, N. A., Singh, V., Deka, S., ... & Mailavaram, R. P. (2021). Tuberculosis: an update on pathophysiology, molecular mechanisms of drug resistance, newer anti-TB drugs, treatment regimens and host-directed therapies. *Current Topics in Medicinal Chemistry*, 21(6), 547-570.
- Canalda-Baltrons, A., Silcocks, M., Hall, M. B., Theys, D., Viberg, L. T., Sherry, N. L., ... & Dunstan, S. J. (2025). Genome graphs reveal the importance of structural variation in *Mycobacterium tuberculosis* evolution and drug resistance. *Nature Communications*, 16(1), 10746.
- Chauhan, M., Barot, R., Yadav, R., Joshi, K., Mirza, S., Chikhale, R., ... & Murumkar, P. R. (2024). The *Mycobacterium tuberculosis* Cell Wall: An Alluring Drug Target for Developing Newer Anti-TB Drugs—A Perspective. *Chemical Biology & Drug Design*, 104(3), e14612.
- Datta, D., Jamwal, S., Jyoti, N., Patnaik, S., & Kumar, D. (2024). Actionable mechanisms of drug tolerance and resistance in *Mycobacterium tuberculosis*. *The FEBS journal*, 291(20), 4433-4452.
- de Araujo, L., Cabibbe, A. M., Mhuulu, L., Ruswa, N., Dreyer, V., Diergaardt, A., ... & Niemann, S. (2023). Implementation of targeted next-generation sequencing for the diagnosis of drug-resistant tuberculosis in low-resource settings: a programmatic model, challenges, and initial outcomes. *Frontiers in public health*, 11, 1204064.
- Dheda, K., Mirzayev, F., Cirillo, D. M., Udhwadia, Z., Dooley, K. E., Chang, K. C., ... & Lange, C. (2024). Multidrug-resistant tuberculosis. *Nature Reviews Disease Primers*, 10(1), 22.
- Dinas Kesehatan Provinsi Sumatera Selatan. Laporan Penanggulangan Tuberkulosis Sumatera Selatan 2024. Published online 2024.
- Dousa, K. M., Kurz, S. G., Bark, C. M., Bonomo, R. A., & Furin, J. J. (2020). Drug-resistant tuberculosis: a glance at progress and global challenges. *Infectious Disease Clinics*, 34(4), 863-886.
- Gengenbacher, M., & Kaufmann, S. H. (2012). *Mycobacterium tuberculosis*: success through dormancy. *FEMS microbiology reviews*, 36(3), 514-532.

- Goletti, D., Meintjes, G., Andrade, B. B., Zumla, A., & Lee, S. S. (2025). Insights from the 2024 WHO global tuberculosis report—more comprehensive action, innovation, and investments required for achieving WHO end TB goals. *International Journal of Infectious Diseases*, 150.
- Gygli, S. M., Borrell, S., Trauner, A., & Gagneux, S. (2017). Antimicrobial resistance in *Mycobacterium tuberculosis*: mechanistic and evolutionary perspectives. *FEMS microbiology reviews*, 41(3), 354-373.
- Jose Vadakunnel, M., Nehru, V. J., Brammacharry, U., Ramachandra, V., Palavesam, S., Muthukumar, A., ... & Soundappan, G. (2025). Impact of *rpoB* gene mutations and Rifampicin-resistance levels on treatment outcomes in Rifampicin-resistant tuberculosis. *BMC Infectious Diseases*, 25(1), 284.
- Katoch, V. M. (2020). Molecular basis of drug resistance in *Mycobacteria*. In *Pathogenicity and Drug Resistance of Human Pathogens: Mechanisms and Novel Approaches* (pp. 3-31). Singapore: Springer Singapore.
- Kemenkes, R. I. Laporan Program Penanggulangan Tuberkulosis Tahun 2022. 2023.
- MacGregor-Fairlie, M., Wilkinson, S., Besra, G. S., & Goldberg Oppenheimer, P. (2020). Tuberculosis diagnostics: overcoming ancient challenges with modern solutions. *Emerging topics in life sciences*, 4(4), 435-448.
- Miotto, P., Cabibbe, A. M., Borroni, E., Degano, M., & Cirillo, D. M. (2018). Role of disputed mutations in the *rpoB* gene in interpretation of automated liquid MGIT culture results for rifampin susceptibility testing of *Mycobacterium tuberculosis*. *Journal of clinical microbiology*, 56(5), 10-1128.
- Nimmo, C., Millard, J., Faulkner, V., Monteserin, J., Pugh, H., & Johnson, E. O. (2022). Evolution of *Mycobacterium tuberculosis* drug resistance in the genomic era. *Frontiers in Cellular and Infection Microbiology*, 12, 954074.
- Pal, R., Bisht, M. K., & Mukhopadhyay, S. (2022). Secretory proteins of *Mycobacterium tuberculosis* and their roles in modulation of host immune responses: focus on therapeutic targets. *The FEBS Journal*, 289(14), 4146-4171.
- Pshennikova, E. S., & Voronina, A. S. (2022). Dormancy: There and back again. *Molecular Biology*, 56(5), 735-755.
- Qadir, M., Faryal, R., Khan, M. T., Khan, S. A., Zhang, S., Li, W., ... & McHugh, T. D. (2024). Phenotype versus genotype discordant rifampicin susceptibility testing in tuberculosis: implications for a diagnostic accuracy. *Microbiology Spectrum*, 12(1), e01631-23.
- Roshdi Maleki, M. (2025). Genetic Analysis of Molecular Mechanisms of Drug Resistance in *Mycobacterium tuberculosis* Against Four Major First-Line Anti-Tuberculosis Drugs (Isoniazid, Rifampin, Ethambutol, and Pyrazinamide). *Infection and Drug Resistance*, 4901-4915.
- Rostamian, M., Kooti, S., Abiri, R., Khazayel, S., Kadivar, S., Borji, S., & Alvandi, A. (2023). Prevalence of *Mycobacterium tuberculosis* mutations associated with isoniazid and rifampicin resistance: a systematic review and meta-analysis. *Journal of Clinical Tuberculosis and Other Mycobacterial Diseases*, 32, 100379.
- Rumende, C. M. (2018). Risk factors for multidrug-resistant tuberculosis. *Acta Medica Indonesiana*, 50(1), 1-1.
- Schami, A., Islam, M. N., Belisle, J. T., & Torrelles, J. B. (2023). Drug-resistant strains of *Mycobacterium tuberculosis*: cell envelope profiles and interactions with the host. *Frontiers in Cellular and Infection Microbiology*, 13, 1274175.
- Singh, R., Dwivedi, S. P., Gaharwar, U. S., Meena, R., Rajamani, P., & Prasad, T. (2020). Recent updates on drug resistance in *Mycobacterium tuberculosis*. *Journal of applied microbiology*, 128(6), 1547-1567.
- Stamilla, A., Recchia, D., Stelitano, G., Maci, L., Marturano, M. C., De Rossi, E., ... & Degiacomi, G. (2025). Uncovering Insights Into the Biology of *Mycobacterium tuberculosis* Using Genetic Tools. *MicrobiologyOpen*, 14(6), e70206.

- Tajer, L., Paillart, J. C., Dib, H., Sabatier, J. M., Fajloun, Z., & Abi Khattar, Z. (2024). Molecular mechanisms of bacterial resistance to antimicrobial peptides in the modern era: an updated review. *Microorganisms*, 12(7), 1259.
- Takenov, N., Kaziyev, A., Mukhamadi, A., Chingissova, L., Toxanbayeva, B., Bismilda, V., ... & Zhunussova, G. (2025). Genetic diversity, drug resistance, and transmission patterns of tuberculosis based on whole-genome sequencing in Almaty, Kazakhstan. *Frontiers in Microbiology*, 16, 1649137.
- Tawfick, M. M., Badawy, M. S. E., Taleb, M. H., & Menofy, N. G. E. (2023). Tuberculosis Diagnosis and Detection of Drug Resistance: A Comprehensive Updated Review. *Journal of Pure & Applied Microbiology*, 17(4).
- Tenchov, R., Hughes, K. J., Ganesan, M., Iyer, K. A., Ralhan, K., Lotti Diaz, L. M., ... & Zhou, Q. A. (2025). Transforming medicine: cutting-edge applications of nanoscale materials in drug delivery. *ACS nano*, 19(4), 4011-4038.
- Tiwari, S., Casey, R., Goulding, C. W., Hingley-Wilson, S., & Jacobs Jr, W. R. (2019). Infect and inject: how *Mycobacterium tuberculosis* exploits its major virulence-associated type VII secretion system, ESX-1. *Microbiology spectrum*, 7(3), 10-1128.
- Wei, X. B., Norsuddin, N. M., Azmi, M. I., & Hamid, H. A. (2026). Artificial Intelligence in Tuberculosis Imaging: A Global Bibliometric Analysis of Research Trends and Collaborations. *Radiography*, 32(3), 103348.
- Xie, Y., Lu, H., Liu, Y., Hu, G., Lian, S., Liu, J., ... & Ding, X. (2025). Unveiling the mechanisms of bacterial resistance and countermeasures. *Pathogens*, 14(11), 1085.
- Xiong, Z., Yang, X., Wang, S., Smart, C. J., Sisson, H. M., Lin, Z., ... & Zhang, B. (2025). Structure and assembly of the MmpL5/MmpS5 efflux transporter from *Mycobacterium tuberculosis*. *Nature communications*, 16(1), 4976.

