

Asian Journal of Research in Zoology

Volume 7, Issue 2, Page 57-63, 2024; Article no.AJRIZ.117221 ISSN: 2582-466X

Predilection Site for the Meat Lover *Trichinella spp* Larvae and its Pathogenesis and Potency in Human Host

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Author's contribution

The sole author designed, analyzed, interpreted and prepared the manuscript.

Article Information

DOI: https://doi.org/10.9734/ajriz/2024/v7i2150

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/117221

Mini-review Article

Received: 10/03/2024 Accepted: 13/05/2024 Published: 15/05/2024

ABSTRACT

Aims: To revisited carefully previously published articles available in the internet regarding predilection site of *Trichinella* spp larvae including their pathogenesis and its potency. **Discussion:** By carefully reviewing previously published literature available in the internet regarding the parasite *Trichinella tropism* to its host's striated muscle, the author briefly withdraw some important keypoint to be revealed. The nematode *Trichinella* spp causes serious zoonosis called trichinellosis, a disease affecting striated or skeletal muscles which consider as one of tropical disease. Even though its natural host varied, but infection among live stocks, such as pigs and other animals, which are raising public health concern. Human infection occurs after consumption of raw or undercooked meat or meat products contain muscle larvae of *T. spiralis*. The tropism of the parasite for individual striated muscles and/or striated muscle groups varies significantly. Trichinella spp. has a direct life cycle where all three life cycle stages (the infective muscle larvae, adult, and new born larvae) happen serially in one host only. Intestine-dwelling

Cite as: Siagian, F. E. (2024). Predilection Site for the Meat Lover Trichinella spp Larvae and its Pathogenesis and Potency in Human Host. Asian Journal of Research in Zoology, 7(2), 57–63. https://doi.org/10.9734/ajriz/2024/v7i2150

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Siagian; Asian J. Res. Zool., vol. 7, no. 2, pp. 57-63, 2024; Article no.AJRIZ.117221

adults of Trichinella produce newborn larvae that bypass the enterocyte, enter the bloodstream and colonize skeletal muscle. The muscle larvae assemble excretory-secretory products which play crucial role in establishing and maintaining persistent parasitism and the host's immune modulation and evasion. It turns out that excretory-secretory products from muscle larvae and mature worm also have hidden medical potential that can be used to treat allergic problems, inflammation-based diseases, autoimmunity and even malignancy.

Conclusion: Trichinellosis is a serious and potentially fatal zoonosis which transmitted through consuming raw or uncooked contaminated meat or its comestibles. Its primary tropism is to the host's striated muscle and infection can persist for a long time facilitated by several reciprocities of its product (e.g., excretory-secretory) with the host's cell and immune system. Fortunately, there are several promising potency in the field of therapeutic and prevention medicine which should be explored intensely.

Keywords: Tropical disease; nematode; tropism; striated muscle; carcasses; trichinosis; trichinelosis.

1. INTRODUCTION

The term tropical diseases encompass all diseases, communicable and no communicable, that occur principally in the tropical countries or tropics, areas that lie between, and alongside, the Tropic of Cancer and Tropic of Capricorn belts [1]. Among those communicable group of disease, neglected helminth infections including trichinellosis or trichinosis, which is still a major health problem [2]. There are eleven known species within the genus Trichinella. These eleven species subdivide into those that invade host muscle cells and encapsulate (surrounded by a collagen capsule) and those that do not encapsulate [3,4]. Trichinella Spiralis, the most common species in this genus, belongs the encapsulated aroup and causes to most human infections and deaths from trichinosis [3]. This genus specifically causing a disease which affecting the host's muscles [1-5].

Even though its natural host varied, but infection among popular live stocks such as pigs [5-8] and other animals, e.g., horses [9], wild game meat (meat from an animal that is typically found in the wild and not raised domestically on a farm for consumption; usually free-roaming mass foragers and hunted for their meat) [10], rats [11], wild birds [12], wild and farmed reptiles [13] etc., which are raising public health concern [14], even though its global burden is much lower than that of other foodborne parasitic diseases (a mean estimated 76 healthy life years lost per billion people per year for human trichinellosis, globally [15].

The aim of this study is to revisited predilection site of Trichinella spp larvae in its host including their pathogenesis and potency along with its comestible implication and effort conducted to prevent transmission.

2. LIFE CYCLE AND PATHOGENESIS

Among many member of helminths which affect human, *Trichinella spp*. are distinctive because it has a direct life cycle [16]; which means that all three life cycle stages of the parasite, namely infective muscle larvae, adult, and new born larvae; Intestine-dwelling adults of Trichinella produce newborn larvae that enter the bloodstream and colonize skeletal muscle [17]. Infection is acquired by consumption of infected raw or undercooked meat or meat based comestibles [3-17].

Under the biochemically pressure of low pH gastric juice, entrapped larvae which is basically anaerobic are released in the host's stomach, followed by the molting process (approximately four times in 30-40-time span) [18,19]. Proteases secreted by Trichinella spiralis intestinal infective larvae directly damage the surrounding junctions of the intestinal epithelial cell monolayer and also arbitrate larval invasion and develop into the adult stage inside the enterocytes of small intestine [20]. The results of study conducted by Song et al [20] stipulate that the parasite enzyme named serine proteases and cysteine proteases play crucial roles in larvae invasion, growth and survival inside the host and that they may be main candidate target molecules for vaccines against larval invasion and development.

After successfully entering enterocyte and become mature, male and female are mating then produce new born larvae [21] are released into circulation and spread throughout the tissues and organs [22] and only those that enter striated muscles mature into muscle larvae [23]. During

the muscular phase, the larvae invade the skeletal muscle fibers inducing a relevant inflammatory reaction aiming for the elimination of the parasite. However, the larvae eventually succeed to build their own home inside the infected myocytes [23]. Muscle invasion results in formation of a capsule surrounding muscle larvae in the region of infected muscles [24]. Once again, this eccentric meat lover Trichinella blessed with the capability to make itself "feel homey like being at home" by way of transforming the infected muscle cell for their own benefit and accomplishing a new type of cell inside the host affected musculature, the so-called nurse cell [25].

The lowest infectious dose of Trichinella larvae is remains unrevealed, but the clinical manifestations of trichinellosis starts to displayed as the number of parasite entering the host increases [26]. Asymptomatic infection could remain silent in human if it is only involving a minimum amount of larvae; Gastrointestinal symptoms manifested as a specific syndrome consist of nausea, diarrhea, vomiting, fatigue, fever, and abdominal discomfort [27], starts very early to develop in case of unintentionally ingestion of hundreds of larvae, perhaps manifest itself clinically within the first 48 hours consuming contaminated meat. after The condition that followed by development of a series of condition which are serious, but scarcely fatal illness [5]. Clinical signs of the disease usually last 4-6 months, rarely longer (up to 2 years).

Chronic form of trichinellosis rarely reported, once in 1983 revealed by two German doctor in their case report regarding biopsies conducted on muscles of five patients with clinical diagnosis chronic neuromuscular disorder. mostly manifested as spinal muscle atrophy. All of them had previous history of acute trichinellosis, the interval between acute parasitic infection and the progressive appearance of the slowly neuromuscular syndrome being of 13 to 26 years respectively. Analysis conducted on the biopsy specimens showed morphological and enzymehistochemical alteration which indicative the presence of progressive neurogenic muscular atrophy. From the Parasite perspective, distinctive encapsulated but still living, enzymepositive parasites were clearly identified with definite signs of focal myositis in the muscle portion surrounding the larva. The possibility pathogenesis correlations between the "chronic" trichinellosis the "degenerative" and

neuromuscular disorder cannot yet be excluded and this still remains to be an unchartered sea of exploration.

In brief, medication option administered in trichinellosis should include potent anthelmintics, alucocorticosteroids. immunomodulating regimen, and combo-preparations which accommodate and compensate protein and electrolyte deficits [27]. Appropriate anthelmintics (type, dosis and length of treatment) are the principal therapy used for trichinellosis. The option of anthelmintics include albendazole. mebendazole and or pyrantel. Rayia et al [23] proposed Bevacizumab as a new option on therapy of muscle phase of Trichinella spiralis infection [28].

3. THE POTENCY

Infective larvae remain alive in striated muscles of the vulnerable host for years [20]; an evidence supported by the study of Sofronic-Milosavljevic et al which revealed the chronic existence of specific antibody responses that still could be recognized even 30 years post primary infection [29]. In case of invasion by Trichinella larvae against the host's immune system, it actually arouses a complex immune response; in human host is better designated by humoral immune response [30] rather than the cellular responses; and this emphasize future prospect of the human host's dynamic humoral response [30] for diagnostic [29] or even vaccine development purposes [31,32] such as reported by Bi et al [32] that revealed the newly identified rTs-ES-1 is potent immunodominant protein secreted by Trichinella stichocytes during natural infection and permits the arousal of fractional protective inimical to immunity in vaccinated mice intentional Trichinella infection. Therefore. findings of this rTs-ES-1 specific protein with the better understanding of its antigenic shiftdvnamicitv [31] is а potential candidate for vaccine development against trichinellosis. In contrast to what happened inside their vulnerable human host, in animals T. spiralis can outstretch a high worm burden without causing prominent clinical symptoms [33].

The initiation of infection depends on first by the annexation of prone intestinal epithelium by infective muscle larvae (ML) and followed secondly with the preservation of parasitism which is marked by the presence of ML in affected muscle cells. The parasite regulatory protein accountable for enzymatic process of these two steps are very important for future investigation [34].

Excretory-secretory products of invading larvae believed to be originate from stichocyte granules in the stichosome, the secretory organelle of the Trichinella's mature muscle larvae [35]. These excretory-secretory products play a pivotal role in parasite's immune evasion and regulation inimical to the host's innate immune system by way of (1) suppressing NET (neutrophil extracellular traps which primary function as a trap for pathogens and facilitating phagocytosis and cytokine production) production and (2) negatively didacte cytokine secretion. The understanding of this excretory-secretory products function for the larvae or worm provides an encouraging area for manufacturing new intervention strategies in other areas of medicine, e.g., in tackling sepsis induced acute lung injury [36] or allergic plethora [29] or autoimmune condition/diseases such as colitis [37] and even malignancies [29].

These important excretory-secretory products engage mainly in the reciprocity with various host cells: firstly, the immune cells, secondly the enterocytes and thirdly the muscle cells, and, through those interaction establishing their role in parasitism and immune response induction, modulation and even evasion [29-32,34-37]. Through these approaches, this nematode generates a perfect milieu for its own suitability and survival in two ways either by modulation of host immune response or affecting host cell gene expression. Extensive exploration of these molecules is important in order to build better understanding regarding (1) the establishment of triumphant parasitism, (2) the development of novel therapies and (3) preventive treatments for inflammatory based disorder.

4. CONCLUSION

Trichinellosis with its related clinical syndrome must always be considered as serious and potentially fatal zoonosis. Transmission occurs through consuming raw or uncooked meat or its comestibles which contaminated with its muscle larvae. Its primary tropism is to the host's striated muscle and can affect the muscle strength and composition in long term. Infection can persist for a long time facilitated by several reciprocities of its product (e.g., excretory-secretory) with the host's organ specific cells (e.g., enterocytes, myocytes) and immune system. Fortunately, there are several promising potency in the field of therapeutic and prevention medicine which should be explored intensely.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- Α. 1. Zumla Α. Ustianowski Tropical diseases: definition. geographic transmission, distribution. and classification. Infect Dis Clin North Am. 2012;26(2):195-205. Available:https://doi.org/10.1016/j.idc.2012 .02.007.
- 2. Hotez PJ, Brindley PJ, Bethony JM, King CH, Pearce EJ, Jacobson J. Helminth infections: the great neglected tropical diseases. J Clin Invest. 2008;118(4):1311-21.

Available:https://doi.org/10.1172/JCI34261

- Pozio E, Zarlenga DS, La Rosa G. The detection of encapsulated and nonencapsulated species of Trichinella suggests the existence of two evolutive lines in the genus. Parasite. 2001;8(2 Suppl):S27-9. Available:https://doi.org/10.1051/parasite/2 00108s2027.
- Furhad S, Bokhari AA. Trichinosis. [Updated 2023 Jul 19]. In: Stat Pearls Treasure Island (FL): Stat Pearls Publishing;2024. Available:https://www.ncbi.nlm.nih.gov/boo ks/NBK536945/
- Kalambhe D, Kaur H, Gill JPS. Trichinella spp. in Slaughtered Pigs of India: From Neglected Disease to an Emerging Food Safety Threat for Public Health. Transboundary and Emerging Diseases. 2024;1-9. Available:https://doi.org/10.1155/2024/755

Available:https://doi.org/10.1155/2024/755 0006.

Gondek M, Knysz P, Pyz-Łukasik R, 6. Łukomska A, Kuriga A, Pomorska-Mól M. Distribution of Trichinella spiralis, Trichinella britovi, and Trichinella pseudospiralis in the Diaphragms and T. spiralis and T. britovi in the Tongues of Experimentally Infected Pigs. Front Vet Sci.2021:8:696284. Available:https://doi.org/10.3389/fvets.202 1.696284.

 Eslahi AV, KarimiPourSaryazdi A, Olfatifar M, de Carvalho LMM, Foroutan M, Karim MR, Badri M, Ketzis JK. Global prevalence of Trichinella in pigs: A systematic review and meta-analysis. Vet Med Sci. 2022;8 (6):2466-2481.

Available:https://doi.org/10.1002/vms3.951
8. Wang N, Bai X, Tang B, Yang Y, Wang X, Zhu H, Luo X, Yan H, Jia H, Liu M, Liu X. Primary characterization of the immune response in pigs infected with Trichinella spiralis. Vet Res. 2020;51(1):17. Available:https://doi.org/10.1186/s13567-020-0741-0.

9. Różvcki M. Korpvsa-Dzirba W. Bełcik A. Bilska-Zaiac Gontarczvk Ε. Α. Kochanowski M. Samorek-Pieróg Μ. Karamon J. Rubiola S. Chiesa F. Cencek T. Validation Parameters of the Magnetic Stirrer Method for Pooled Sample Digestion for Trichinella spp. in Horse Meat Based on Proficiency Tests Results. Int J Environ Res Public Health. 2022; 19(21):14356.

https://doi.org/10.3390/ijerph192114356.

- McIntyre L, Pollock SL, Fyfe M, Gajadhar A, Isaac-Renton J, Fung J, Morshed M. Trichinellosis from consumption of wild game meat. CMAJ. 2007;176(4):449-51. Available:https://doi.org/10.1503/cmaj.061 530.
- Bilska-Zając E, Różycki M, Antolak E, Bełcik A, Grądziel - Krukowska K, Karamon J, et al. Occurrence of Trichinella spp. in rats on pig farms. Ann Agric Environ Med. 2018;25(4):698-700. Available:https://doi.org/10.26444/aaem/99 555
- Rugna G, Marucci G, Bassi P, Gelmini L, D'Annunzio G, Torreggiani C, et al. Trichinella surveillance program in wild birds, Emilia-Romagna (northern Italy), 2006–2021. First report of Trichinella pseudospiralis in western marsh harrier (Circus aeruginosus) in Italy. International Journal for Parasitology: Parasites and Wildlife.2022;19:191-195, Available:https://doi.org/10.1016/j.ijppaw.2 022.09.006.
- Pozio E, Foggin CM, Gelanew T, Marucci G, Hailu A, Rossi P, Morales MA. Trichinella zimbabwensis in wild reptiles of Zimbabwe and Mozambique and farmed reptiles of Ethiopia. Vet Parasitol. 2007;143(3-4):305-10.

Available:https://doi.org/10.1016/j.vetpar.2 006.08.029.

- Yayeh M, Yadesa G, Erara M, Fantahun S, Gebru, Birhan M Epidemiology, diagnosis and public health importance of Trichinellosis. Online J. Anim. Feed Res. 2020;10(3):131-139. Available:https://dx.doi.org/10.36380/scil.2 020.ojafr18
- Devleesschauwer B, Praet N, Speybroeck N, Torgerson PR, Haagsma JA, De Smet K, Murrell KD, Pozio E, Dorny P. The low global burden of trichinellosis: evidence and implications. Int J Parasitol. 2015; 45(2-3):95-9. Available:https://doi.org/10.1016/j.ijpara.20 14.05.006
- Gamble HR. Trichinella. Ed(s): Batt CA, Tortorello ML. Encyclopedia of Food Microbiology (Second Edition), Academic Press.2014;638-643, ISBN 9780123847331. Available:https://doi.org/10.1016/B978-0-12-384730-0.00336-0.
- Lee JL, Rosenberg HF (eds). Chapter 10 -Eosinophils: Mediators of Host-Parasite Interactions. In books Eosinophils in Health and Disease, Academic Press. 2013;301-327. ISBN 9780123943859.

Available:https://doi.org/10.1016/B978-0-12-394385-9.00010-9.

- Mostafa EM, Atwa HA. Intestinal mastocytosis in Trichinella spiralis infection: immunohistochemical study in murine model. Parasitologists United Journal. 2020;13(1):52-59. Available:https://doi.org/10.21608/puj.2020 .24540.1060
- Gagliardo LF, McVay CS, Appleton JA. Molting, ecdysis, and reproduction of Trichinella spiralis are supported in vitro by intestinal epithelial cells. Infect Immun. 2002;70(4):1853-9. Available:https://doi.org/10.1128/IAI.70.4.1 853-1859.2002.
- 20. Song YY, Lu QQ, Han LL, Yan SW, Zhang XZ, Liu RD, Long SR, Cui J, Wang ZQ. Proteases secreted by Trichinella spiralis intestinal infective larvae damage the junctions of the intestinal epithelial cell monolayer and mediate larval invasion. Vet Res. 2022;53(1):19.

Available:https://doi.org/10.1186/s13567-022-01032-1.

- 21. Gardiner CH. Habitat and reproductive behavior of Trichinella spiralis. J Parasitol. 1976;62(6):865-70.
- Stewart GL, Despommier DD, Burnham J, Raines KM. Trichinella spiralis: behavior, structure, and biochemistry of larvae following exposure to components of the host enteric environment. Exp Parasitol. 1987;63(2):195-204. Available:https://doi.org/10.1016/0014-4894(87)90162-7.
- Rayia DA, Othman A, Harras S, Helal D, Dawood L, Soliman S. Bevacizumab: A new take on therapy of muscle phase of Trichinella spiralis infection. Acta Trop. 2022;230:106409. Available:https://doi.org/10.1016/j.actatropi ca.2022.106409.
- Wu Z, Sofronic-Milosavljevic Lj, Nagano I, Takahashi Y. Trichinella spiralis: nurse cell formation with emphasis on analogy to muscle cell repair. Parasit Vectors. 2008; 1(1):27. Available:https://doi.org/10.1186/1756-
- 3305-1-27.25. Despommier DD. How does Trichinella spiralis make itself at
- Trichinella spiralis make itself at home? Parasitol Today. 1998;14(8):318-23.

Available:https://doi.org/10.1016/s0169-4758(98)01287-3.

 Diaz JH, Warren RJ, Oster MJ. The Disease Ecology, Epidemiology, Clinical Manifestations, and Management of Trichinellosis Linked to Consumption of Wild Animal Meat. Wilderness & Environmental Medicine. 2020;31(2):235-244.

Available:https://doi.org/10.1016/j.wem.20 19.12.003

- 27. Kociecka W. Trichinellosis: human disease, diagnosis and treatment. Vet Parasitol. 2000;93(3-4):365-83. Available:https://doi.org/10.1016/s0304-4017(00)00352-6.
- Gullotta F, Fröscher W. Chronische Trichinose und neuromuskuläre Erkrankungen. Morphologische und pathogenetische Aspekte [Chronic trichinosis and neuromuscular diseases. Morphologic and pathogenesis aspects]. Arch Psychiatr Nervenkr. 1983;232(6):479-87.

Available:https://doi.org/10.1007/BF00344 062.

- 29. Sofronic-Milosavljevic L, Ilic N, Pinelli E, Gruden-Movsesijan A. Secretory Products of Trichinella spiralis Muscle Larvae and Immunomodulation: Implication for Autoimmune Diseases, Allergies, and Malignancies. J Immunol Res. 2015;2015: 523875. https://doi.org/10.1155/2015/523875.
- 30. Alcántara P, Correa D. Human humoral immune responses against Trichinella spiralis. Int J Parasitol. 1993;23(5):657-60. Available:https://doi.org/10.1016/0020-7519(93)90173-v.
- Boireau P, Vallée I, Karajian G, Wang X, Liu M. Chapter 16 - Antigenic shift during Trichinella cycle, consequences for vaccine developments, Editor(s): Fabrizio Bruschi, Trichinella and Trichinellosis. Academic Press. 2021;455-516, ISBN 9780128212097. Available:https://doi.org/10.1016/B978-0-12-821209-7.00014-7.
- Bi K, Yang J, Wang L, Gu Y, Zhan B, Zhu X. Partially Protective Immunity Induced by a 20 kDa Protein Secreted by Trichinella spiralis Stichocytes. PLoS One. 2015;10(8):e0136189. Available:https://doi.org/10.1371/journal.po ne.0136189.
- Ribicich M, Pasqualetti MI, Fariña FA. Chapter 9 - Trichinellosis in animals. Ed(s): Bruschi F. Trichinella and Trichinellosis. Academic Press. 2021;315-331, ISBN 9780128212097. Available:https://doi.org/10.1016/B978-0-12-821209-7.00013-5.
- Hao HN, Song YY, Ma KN, Wang BN, Long SR, Liu RD, Zhang X, Wang ZQ, Cui J. A novel C-type lectin from Trichinella spiralis mediates larval invasion of host intestinal epithelial cells. Vet Res. 2022;53 (1):85. Available:https://doi.org/10.1186/s13567-

022-01104-2. Despommier DD, Müller M. The

- 35. Despommier DD, Müller M. The stichosome and its secretion granules in the mature muscle larva of Trichinella spiralis. J Parasitol. 1976;62(5):775-85.
- 36. Li H, Qiu D, Yang H, Yuan Y, Wu L, Chu L, Zhan B, Wang X, Sun Y, Xu W, Yang X. Therapeutic Efficacy of Excretory-Secretory Products of Trichinella spiralis Adult Worms on Sepsis-Induced Acute Lung Injury in a Mouse Model. Front Cell Infect Microbiol. 2021;11:653843.

Siagian; Asian J. Res. Zool., vol. 7, no. 2, pp. 57-63, 2024; Article no.AJRIZ.117221

Available:https://doi.org/10.3389/fcimb.202 1.653843.

37. Yang X, Yang Y, Wang Y, Zhan B, Gu Y, Cheng Y, Zhu X. Excretory/secretory products from Trichinella spiralis adult worms ameliorate DSS-induced colitis in mice. PLoS One. 2014;9(5): e96454.

Available:https://doi.org/10.1371/journal.po ne.0096454.

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