TurnitinBioactivecompoundsan tidiabeticandantimicrobialpote ntialofpinangseeds

by Edi Wibowo

Submission date: 15-Nov-2023 03:53PM (UTC+0700)

Submission ID: 2227750995

File name: 2060-9156-1-PB.pdf (419.47K)

Word count: 5969

Character count: 34350

JURNAL AISYAH: JURNAL ILMU KESEHATAN, 8(3) 2023

Available online at: https://aisyah.journalpress.id/index.php/jika/
DOI: 10.30604/jika.v8i3.2060

Bioactive compounds, antidiabetic and antimicrobial potential of pinang seeds extract (Areca catechu l.)

Trini Suryowati^{1*)}, Forman Erwin Siagian², Hera Maheswari³, Yusias Hikmat Diani⁴, Rini Anjarwati Kusuma⁵

Department of Biochemistry, Faculty of Medicine, Universitas Kristen Indonesia

Department of Parasitology, Faculty of Medicine, Universitas Kristen Indonesia

Division Physiology, School of Veterinary Medicine and Biomedical Sciences, Institut Pertanian Bogor

⁴Department of Continuity Medicine, Faculty of Medicine, Universitas Kristen Indonesia, ⁵Medical Student, Faculty of Medicine, Universitas Kristen Indonesia



Diabetes mellitus is a chronic metabolic disorder affecting people of all ages. The critical aspect of fruits is that rich sources of antioxidants may act in combination with each other phytochemicals to provide their protective effect. Pinang (Areca catechu L.) fruit is edible as the local indigenous plant from West Irian Jaya (Papua) Indonesia. This study was aimed at investigating the biologically active compounds of seeds, fruits, and leaves, the α -glucosidase inhibitory and antibacterial activity of seeds of Pinang against Gram-positive bacteria Staphylococcus aureus ATCC 25923, and Gramnegative bacteria Escherichia coli ATCC 25922. Pinang fruits were extracted by using the maceration method and ethanol solvent. Identify the chemical compounds in seeds by GC-M 10 echnique, test to a-glycosidase inhibitory effect was measured with spectrophotometric. Well, the diffusion method was employed in evaluating the antimicrobial property of extracts. The evaluation of the bioactive compound of Pinang fruits revealed the presence of Vitamin E (0.20%). The inhibition of α -glucosidase of seeds extract of ICs0 values was 82.74 ppm, and the global standard w 11.34 ppm. This study confirmed that Pinang seeds contain glucosidase activity that has the potency to inhibit glucose. The antimicrobial activity was performed against bacteria as it showed zone inhibition. These results indicated that Pinang seed extracts exerted potent inhibitory effects against α -glucosidase and inhibited the proliferation of Gram-negative microorganisms.

Keywords: α-glucosidase, antimicrobial, Pinang (Areca catechu L.) extract.

ABSTRAK

Diabetes melitus merupakan kelainan metabolisme kronis yang menyerang semua usia. Aspek penting dari buah-buahan adalah sumber antioksidannya yang kaya dapat bekerja bersama dengan fitokimia lainnya untuk memberikan efek perlindungan. Buah Pinang ($Areca\ catechu\ L$.) dapat dimakan sebagai tanaman asli lokal dari Irian Jaya Barat (Papua) Indonesia. Penelitian ini bertujuan untuk mengetahui senyawa aktif biologis biji, buah, dan daun, daya hambat α -glukosidase dan aktivitas antibakteri biji Pinang terhadap bakteri Gram positif Staphylococcus aureus ATCC 25923, dan bakteri Gram negatif Escherichia coli ATCC 25922. Buah pinang diekstraksi menggunakan metode maserasi dan pelarut etanol. Identifikasi senyawa kimia dalam biji dengan teknik GC-MS, uji efek penghambatan α -glikosidase diukur dengan spektrofotometri. Nah, metode difusi digunakan dalam mengevaluasi sifat antimikroba dari ekstrak. Evaluasi senyawa bioaktif buah Pinang menunjukkan adanya Vitamin E (0,20%). Daya hambat α -glukosidase ekstrak biji kakao nilai IC50 sebesar 82,74 ppm, dan standar global sebesar 0,34 ppm. Penelitian ini menegaskan bahwa biji Pinang mengandung aktivitas glukosidase yang berpotensi menghambat glukosa. Aktivitas antimikroba dilakukan terhadap bakteri karena menunjukkan zona penghambatan. Hasil ini menunjukkan bahwa ekstrak biji Pinang memberikan efek penghambatan yang kuat terhadap α -glukosidase dan menghambat perkembangbiakan mikroorganisme Gram-negatif.

Kata Kunci: α-glukosidase, antimikroba, ekstrak Pinang (Areca catechu L.).

Corresponding author: Trini Suryowati

Department of Biochemistry, Faculty of Medicine, Universitas Kristen Indonesia

Jl. Mayjen Sutoyo No. 2, East Jakarta 13630, Jakarta, Indonesia

Email: trini.suryowati@uki.ac.id

INTRODUCTION

Diabetes mellitus (DM) is a metabolic disorder in which a person has abnormally high blood glucose levels. It is a severe, significant chronic with potency life-threatening, in which the pancreas hormone improperly regulates the homeostasis of carbohydrates, which impacts the health status. Blood glucose control is critical for preventing or reversing diabetic complication (Doan et al., 2018; Demir et al., 2021). At least estimated in 2019 that 463 million people have diabete, projected to reach 578 million by 2030 and 700 million by 2045 (IDF 2019).

In recent years, there has been a resurge 4 e of interest in treating diabetes mellitus (DM) using herbal remedies, mainly because of their non-toxic nature. The World Health Organization has also recommended the assessment of the effectiveness of herbal plants, particularly in cases where safe and modern pharmaceuticals are lacking. Exploring bioactive compounds in plants with potential antidiabetic properties is of utmost importance. It is well known that the presence and effectiveness of these bioactive compounds in plants can vary depending on their location. Many plant species have been documented as having hypoglycemia properties and are traditionally used for preventing and managing diabetes. Despite Sefforts to develop hypoglycemia agents from both natural and synthetic sources, diabetes and its associated complications continue to pose significant medical challenges (Arya et al., 2015; Chen H, 2021; Ansari et al., 2022). Numerous medicinal plants and their products, including active principles and crude extracts, have been reported in the literature for their traditional use in diabetes control (Hegde et al., 2016; Saad B, Kmail A, Haq SZ, 2022; Nabi M, Tabassum N, Ganai BA, 2022). Our ongoing investigation focused on assessing the inhib ary effects of Pinang (Areca catechu L.) plants on the α-glucosidase enzyme. These enzymes play crucial roles in various biochemical processes related to metabolic disorders and diseases, including diabetes. As such, there has been a concerted effort to design efficient α-glucosidase inhibitors with promising potential applications (Chipiti et al. 2015; Di Santo MC, D'Antoni CL, Rubio AP, Alaimo A, Pérez OE, 2021).

Regenerate



Figure 1. Pinang Fruits (Areca Catechu L.)

Alpha-glucosidase inhibitors are carbohydrates that function as competitive antagonists for the enzymes necessary for carbohydrate digestion, specifically the α -glucosidase enzymes located in the brush border of the small intestine. These intestinal α -glucosidase enzymes break down oligosaccharides, trisaccharides, and disaccharides into glucose and other monosaccharides within the small intestine. Inhibitin 14 ese enzyme systems reduces the rate at which carbohydrates are digested, leading to decreased glucose absorption, as the carbohydrates are not broken down into glucose molecules. The short-term impact of these drug therapies in diabetic patients is a reduction in current blood glucose levels, and the long-term effect is a slight decrease in hemoglobin levels (Hussain et al. 2019: Jeong D, Priefer R, 2022; Schauer et al., 2017).

Plants produce secondary metabolites in the form of phenolic compounds, also known as polyphenols, which are known for their antioxidant properties and, as a result, offer beneficial physiological effects. Polyphenols are among the most common antioxidant phytochemicals, and they are recognized for their ability to quench singlet oxygen and scavenge free radicals, thereby retarding the oxidation of lipids. Previous research has indicated that Areca catechu L. fruits are rich in phenolics and tannins, although their antioxidant activities remain relatively unexplored. In traditional markets in Papua, Pinang seeds are frequently used to prepare beverages like coffee.

Extensive research has been conducted on the antimicrobial properties of specific plant species. The pharmacological properties of phytochemicals suggest their potential for antimicrobial and antifungal effects (Shah et al., 2018). The mechanism through which antioxidants act against bacteria involves damaging the integrity of the cell wall and cell membrane, inhibiting intracellular enzyme activity, increasing the levels of reactive oxygen species (ROS), influencing the expression of associated genes, ultimately leading to bacterial apoptosis (Hu et al., 2019; Zhang et al., 2020; Ren X, An P, Zhai X, Wang S, Kong Q, 2019). These findings offer valuable insights into identifying bioactive compounds within Pinang seeds (Areca catechu L.) using GCMS, and the inhibitory effect on α-glucosidase activity was quantified using spectrophotometry. Furthermore, the research suggests a potential antimicrobial effect against both Gram-positive

Jurnal Aisyah: Jurnal Ilmu Kesehatan, 8 (3) 2023, - 1243

Staphylococcus aureus ATCC 25923 and Gram-negative Escherichia coli ATCC 25922, as evidenced by the observed zones of inhibition. This study holds the promise of enhancing the utility of Pinang fruits across various domains, with a particular emphasis on their applications within medicine.

RESEARCH METHOD

Plant Extraction

Dried and fully matured fruits of Areca catechu L., commonly known as Pinang, were sourced from a traditional market in Papua, Indonesia. The extraction process involved soaking the sample in ethanol for a period of 48 hours, followed by subsequent extraction. Chloroform was employed for the extraction process, yielding a chloroform-soluble extract, which was then subjected to centrifugation at 10,000 rpm for a duration of 20 minutes. The resulting clear supernatant oil was subsequently analyzed using GCMS.

Procedures for Gas Chromatography-Mass Spectrometry (GC-MS)

Chemical compound identification was carried out using the SHIMAZU Gas Chromatography 5890-11 ft 3n Japan, equipped with a fused GC column composed of polymethyl silicon, specifically OV 101, with dimensions of 0.25 mm x 50 m. The temperature programming ranged from 80°C to 200°C, with an initial hold at 80°C for 1 minute, followed by a rate of increase at 5°C/min, and a final hold at 200°C for 20 minutes. The Flame Ionization Detector (FID) was set at 300°C, while the injection temperature was maintained at 250°C. Nitrogen served as the carrier gas, flowing at a rate of 1 cm³/min, with a split ratio 61:75. For mass spectrum analysis, a GCMS-QP 2010 Plus instrument from Shimazu Japan was used, featuring an injector temperature of 230°C and a carrier gas pressure of 100 kph. The column had a length of 30 m, a diameter of 0.25 mm, and a flow rate of 50 m/min. The eluents were automa 3 cally directed into the Mass Spectrometer, where the detector voltage was set at 1.5 kv, and data were collected at a sampling rate of 0.2 seconds. The Mass Spectrometa was also equipped with a computer-fed Mass Spectra data bank. The centrifuge employed in the study was the HERMCE Z 233 M-Z from Germany. Reagents and solvents, such as ethanol for analytics, were sourced from Merck Germany (Iwu et al., 2018; Shahvar A, Shamsaei D, Saraji M, 2020; Kachangoon R, Vichapong J, Santaladchaiyakit Y, Srijaranai S, 2020).

Assessment of a-Glucosidase Inhibition

The initial inhibition test involved a reaction mixture comprised of $50 \mu L$ of 0.1 M Phosphate buffer (pH: 7.0), 25 μL of 0.5 mM 4-nitrophenyl α -D-glucopyranoside, 10 μL of the sample with a concentration of $500 \mu g$ mL-1, and 25 μL of α -glucosidase solution. This reagent was incubated at 37°C for 11 ninutes, and the enzymatic reaction was terminated by adding $100 \mu L$ of 0.2 M Na2CO3 (200 mM). To assess the enzymatic absorbance of each mixture, a spectrophotometric measurement was performed using a microplate reader, recording the absorbance at 400 nm.

The negative control involved the reaction between the substrate and enzyme without the presence of an inhibitor. Simultaneously, a blank solution represented the reaction system where the enzyme and inhibitor were absent. This set of actions was performed in triplicate for accuracy. Acarbose was dissolved in a solution containing buffer and 2 N HCL in a 1:1 ratio and was employed as a positive control, serving as an α -glucosidase inhibitor. The calculation of % inhibition was as follows:

The formula for calculating the inhibition percentage is expressed as: % inhibition = [1 - (sample absorbance / control absorbance)] x 100%. The sample concentration and its corresponding percentage of inhibition were then plotted on a graph using the equation Y = a + bx. The term "inhibition activity value" (IC50) was 2 efined as the concentration at which the sample could inhibit 50% of the enzyme's activity. The assessment of enzyme inhibition activities for the α -glucosidase assay was conducted with slight adjustments, as per the work of Yin et al. in 2014.

Determination of Antibacterial Activity

The antibacterial properties of the Pinang extract were assessed through the agar well diffusion method. Escherichia coli ATCC 25922 (Gram-negative) and Staphylococcus aureus ATCC 25923 (Gram-positive) bacteria strains were utilized to evaluate the antibacterial efficacy of these extracts. The extract solution was prepared by dissolving 0.1 g of the extract in 100 mL of solvent, consisting of a mixture of distilled water and absolute ethanol, resulting in a 100 mg/mL concentration. This solution was further diluted to achieve 5%, 10%, and 15% concentrations in distilled water. Subsequently, 25 µL of the extract solution was added to prepared discs measuring 6 mm in diameter, which were placed on Muller-Hilton agar as part of the sensitivity test, followed by the determination of the minimum inhibitory concentration. Chloramphenicol was employed as the positive control, and the inhibition zone produced by the extract was measured after 24 hours of incubation at 37°C (Arvind et al., 2014; Dan et al. Q, Zhang F, Wang WW, Gao JM, 2019; Ha KS, Jo SH, Mannam V, Kwon YI, Apostolidis E, 2016).

RESULTS AND DISCUSSION

The Results of Phytochemical Screening of Pinang (Areca catechu L) Plant

Analytical Gas Chromatography-Mass Spectrometry 7 iC-MS) was employed to determine the phytochemical components within the ethanol extracts of Pinang seeds. The identification of these compounds was confirmed by examining the molecular formula, retention time, and peak area, as illustrated in Figure 2 and detailed in Table 1.

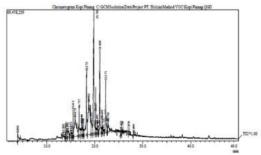


Figure 2: The components in Pinang seeds

There are 30 components metabolites in the results of Pinang seeds extract, using GCMS in Table 1

Table 1 GC-MS analysis of an extract of Pinang seeds

Peak	Retention Time (s)	Quality	Conc (%)	Name of the compound		
1	3.893	48498945	1.49	Ethanol (CAS) Ethyl alcohol		
2	13.363	13657573	0.42	The compound with the CAS identifier Oxiranecarboxamide, 2-ethyl-3-propyl-		
				can be more simply referred to as Oxanamide.		
3	13.864	24662469	0.76	1 enol, 2-methoxy- (CAS) Guaiacol		
4	14.117	44937130	1.38	9 (1H-IMIDAZOL-4-YL)-PROPAN-1-OL		
5	14.372	60537478	1.86	9-Oxabicyclo [6.1.0] nonan-4-one (CAS)4-Oxo-9-oxabicyclo [6.1.0] nonane		
6	14.972	39618113	1.22	2-Noreplagiodial-D4		
7	15.217	23188034	0.71	1-D1-CIS-1,2-CYCLOHEXANEDIOL		
8	15.442	62348894	1.91	2-Propanol, 1-amino- (CAS) 1-Amino-2-propanol		
9	15.871	3105816773	9.38	NICKEL1-AMINO-1,9-DIISOTHIOCIPNO-4,8-DI-AZAUNDECAMINE		
10	16.717	1708952377	3.34	Benzenepropanenitrile. betaimino- (CAS) Acetonitrile, benzimidoyl-		
11	17.240	96641382	2.97	2,4-Pentanediamine, 2-methyl- (CAS) 2,4-DIAMINO-2-METHYLPENTANE		
12	17442	31681968	0.97	2-(5-BROMO-PENT-3-YNYLOXY)-TETRAHYDRO-PYRAN		
13	17.542	37309950	1.14	Ethanamine, N-ethyl-N-hydroxy- (CAS) N, N-Diethylhydroxyamine		
14	18.273	16832467	9.72	1,4-Butanediol (CAS) Sucol B		
15	18.735	322898581	16.04	Tetrahydro-4H-pyran-4-ol		
16	19.291	219039910	3.65	1,2,3,4-Butanetetrol, [S-(R*,R*)]- (CAS) L-ERYTHRITOL		
17	19.785	144245636	16.70	Piperazine, 2-methyl- (CAS) 2-Methylpiperazine		
18	20.167	7 5813436	2.33	3(2H)-Furanone, dihydro-5-methyl- (CAS) 5-METHYL-3-OXO- TETRAHYDROFURAN		
19	20.491	49335566	1.51	2H-Pyranol, tetrahydro-3(or 5)-methyl- (CAS) 3-METHYL HYDROXY TETRAHYDRO PYRAN		
20	20.717	12642485	0.39	5-IYDROXY-4-NITROBENZOTRIAZOLE		
21	21.038	571995947	7.89	6H-Pyrazolo[1,2-a][1,2,4,5]tetrazine, hexahydro-2,3-dimethyl- (CAS) 1,3,4,6- TETRAAZABICYCLO		
22	21.492	42383339	1.30	1,4-Butanediol (CAS) Sucol B		
23	21.715	30315579	0.93	Ethanamine, N-ethyl-N-hydroxy- (CAS) N, N-Diethylhydroxyamine		
24	22.272	102784108	6.22	13-Oxabicyclo[10.1.0]tridecane (CAS) Epoxycyclododecane		
25	22.567	24474276	0.75	1-(CYCLOHEXYL-HYDROXY-AMINO)-PROPANE-2-OL		
26	22.755	70840139	2.17	26 22.755 7 0840 139 2.7 Ethanol, 2,2',2"-nitrilotris- (CAS) Triethanolamine		
27	25.422	22301874	0.68	1-Piperazinecarboxylic acid, ethyl ester (CAS) Ethyl N-piperazinocarboxylate 3259080273 100.00		
28	25.750	24595061	0.75	3-METHYLCYCLOPENTANONE-2.2.5.5-D4		
29	27.070	30551180	0.94	1,3-Propanediamine, N, N-diethyl- (CAS) 3-DIETHYLAMINO PROPYL		
	_,,,,,,			AMINE		
30	28.099	14975956	0.46	Piperazine, 2-methyl- (CAS) 2-Methylpiperazine		
		3259080273	100.00			

Jurnal Aisyah: Jurnal Ilmu Kesehatan, 8 (3) 2023, - 1245

The observed of the ethanol extract Pinang fruits, there were 50 compounds identified. The identification was assured by observing the molecular formula, retention time, and peak area of the data shown in Figure 3 and Table 2.

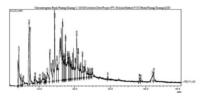


Figure 3. The components in Pinang fruits

There are 50 component metabolites in the results of Pinang fruits, using GC-MS in Table 2.

Table 2 GC-MS analysis of an extract of Pinang fruits

Peak	Retention Quality Conc Name of the comp		Name of the compound		
1	3.993	4 1267771	1.84	Acetic acid, hydroxy-, methyl ester (CAS), Methyl glycolate 5 5.466 1 6964033 0.75 Cyclopropyl carbinol	
2	4.247	9 0587839	4.03	Acetic acid, hydroxy-, methyl ester (CAS), Methyl glycolate	
3	4.792	1 7187970	0.76	(Methylcyclopentadienyl)	
				[(2,4,6-triisopropylphenyl) phosphane] molybdenum tetrachloride	
4	5.167	43166690	0.193	[2,3,5,6-tetrafluoro-4-trifluoromethyl) phenoxy] pregna-3,5-diene-20-one	
5	5.466	1 6964033	0.75	5 5.466 1 6964033 0.75 Cyclopropyl carbinol	
6	7.114	1 1651785	4.97	2-DEUTERO-2-PROPANOL	
7	7.392	4 7463832	2.11	Oxirane (CAS) Epoxyethane	
8	8.617	8 7443009	0.33	Chlorine oxide (Cl21) (CAS) Chlorine monoxide	
9	8.802	2 3545163	1.05	3-OXA-BICYCLO [3.2.0] HEPT-6-ENE-2,4-DIONE	
10	9.917	5957835	0.27	1,4-Butanediol (CAS) Sucol B	
11	10.150	9309750	0.41	3-Butyn-1-ol (CAS) 3-Butynol	
12	10.994	24010146	1.07	L-(-)-ASPARAGIN	
13	11.392	5481624	0.24	Hexatriacontane (CAS) n-Hexatriacontane	
14	11.617	2727468	0.12	O13 2-YNOIC ACID	
15	11.842	3723801	0.17	2,3,6,7-tetramethyl-9,10-bis(4-methylphenylsulfonyloxy)-1,4,4, alpha.,5,8,8a, beta.,9	
				12ta.,9a. beta	
16	12.226	4 9111091	2.19	6-DIMETHYLAMINO-3,5-DIHYDROXY-1,2,4-TRIAZINE	
17	12.96 1	42 2142946	5.44	1H-Pyrrole (CAS) Pyrrole	
18	13.831	4 8725150	2.17	Phenol, 2-methoxy- (CAS) Guaiacol	
19	14.321	554134633	6.86	METHYL-OXIRANE-2-CARBOXYLIC ACID	
20	14.993	6 4642977	2.88	2-(3.5-DIMETHYL-1-PYRAZOLYL)SUCCINIC ACID	
21	15.922	895078510	13.13	15.92 289 5078510 13. 9 TRANS- 3,5-DEUTERON HYDROXY CYCLOPENTENE	
22	16.570	6 8506413	3.05	16.570 6 8506413 3.05 Phenol, 2,6-dimethoxy- (CAS) 2,6-Dimethoxyphenol	
23	16.800	3 6385216	1.62	Tentanal (CAS) n-Pentanal	
24	16.992	4 1792593	1.86	HYDRAZINECARBOXAMIDE, 2-(2,6-CYCLOOCTADIEN-1-YLIDENE)-	
25	17.167	3 7535112	1.67	HEXA-2.4-DIYNE-1.6-DIOL	
26	17.407	5 4810399	2.44	5-ETHYL-2-METHYL-PYRIDIN-4-YLAMINE	
27	17.99	121 7652166		17.99 121 7652166 5.24 1,2-(1'-METHYLTRIMETHYLENE)-DIBORANE	
28	18.319	5 0954593	2.27	13.19 5 0954593 2.27 Ethanone, 1-(4-pyridinyl)-, oxime	
29	18.615	3 4782251	1.55	N-[(5-METHYL-2-PYRIDYL)AMINOMETHYL]-4-NITROPHTHALIMIDE	
30	18.792	1 7844319	0.79	Hexanedinitrile (CAS) Adiponitril	
31	18.967	45080280	2.01	1-Propyne, 3,3'-oxy bis- (CAS) Dipropargyl ether	
32	19.442	88899516	3.96	19.442 88899516 3.96 3-[(Z)-1-Butenyl]-4-vinyl cyclopentene Peak# R.Time Area	
32	17.772	00099310	3.90	Conc%PUSLITBANG HASIL HUTAN REPORT Name	
33	20.131	41781363	1.86	2-Piperidinone, 6-methyl- (CAS) 6-Methyl-2-piperidone	
34	20.518	128857593	5.73	5-ETHYL-N-METHYL-3-BUTENYLAMINE	
35	20.934	57932585	2.58		
				9 TRAAZABICYCLO	
36	21.464	15094044	0.67	7-Oxabicyclo[4.1.0]heptane(CAS) Cyclohexene oxide	
37	21.667	31817404	1.42	5,5-D2-TRANS-3,4-DIHYDROXY-CYCLOPENTENE	
38	22.167	48545178	2.16	Propenal dimethylhydrazine	
39	22.492	12972899	0.58	5-Thiomethylfurfural	
40	22.912	53046873	2.36	FORMAMIDE, N,N'-1,3-PROPANEDIYLBIS-	
41	23.669	19268539	0.86	TRANS- 3,5-DIDEUTERO HYDROXY CYCLOPENTENE	
42	23.942	8284749	0.37	1,3,2-Dioxaphosphorinane, 2,5,5-triphenyl-, 2-oxide (CAS) 2-OXO-2,5,5-TRIPHENYL-1,3,2-DIOXAPHOSPHORINANE	

Jurnal Aisyah: Jurnal Ilmu Kesehatan, 8(3) 2023, - 1246

43	24.317	11892832	0.53	-Pentanol, 2-amino-4-methyl-, (S)-
44	24.767	7970280	0.35	O, N-PERMETHYLATED AC-ALA-HIS
45	24.956	13517553	0.60	Oxiranecarboxamide, 2-ethyl-3-propyl- (CAS) Oxanamide
46	29.261	1575985	0.07	1-(2-METHYL-ALLYL)-AZETIDINE 2247262259 100.00
47	38.208	4864971	0.22	Urea, N,N'-diethyl- (CAS) N,N'-Diethylurea
48	38.602	4453877	0.20	Vitamin E
49	41.917	23707032	1.05	Heptanal (CAS) n-Heptanal
50	42.262	21961642	0.98	Piperazine, 2-methyl- (CAS) 2-Methylpiperazine
		2247262259	100.00	

The identified of the ethanol extract Pinang leaves, there were 50 compounds identified. The observed was assured by observing the molecular formula, retention time, and peak area of the data shown in Figure 4 and Table 3.

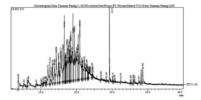


Figure 4. The components in Pinang leaves

There are 60 components metabolites in the results of Pinang leaves extract, using GCMS in Table 3:

Table 3. GC-MS analysis of an extract of Pinang leaves

Peak	Retention	Quality	Conc	Name of the compound
	Time (s)		(%)	•
1	3.650	27475350	1.09	(3S,4S)-3,4-Bis(methoxy ethoxy)pyrrolidine
2	5.213	10234497	0.41	Propanenitrile, 3-(methylamino)-
3	6.167	11332584	0.45	45 N-BUTANE-1,1,1-D3
4	6.469	26579943	1.05	Cyclobutanol (CAS) Cyclobutyl hydroxide
5	8.667	5856399	0.23	Cyclobutanol (CAS) Cyclobutyl hydroxide
6	10.092	6580817	0.26	2-Pentenenitrile (CAS) 1-Cyano-1-butene
7	10.626	6532023	0.26	Formamide, N-(2-methylpropyl)- (CAS) N-Isobutylformamide
8	11.102	11998516	0.48	Ethanamine, N, N-difluoro- (CAS) ETHYL-N, N-DIFLUOROAMINE
9	11.492	6453835	0.26	Lup-20(29)-en-21-ol, 3,28-bis[(tetrahydro-2H-pyran-2-yl)oxy]-,3,3-dimethyl butanoate
				(3.beta.,21.10
10	11.971	61766179	2.45	6-DIMETHYLAMINO-3,5-DIHYDROXY-1,2,4-TRIAZINE
11	12.512	44464290	1.76	2,6-Nonadien-1-ol (CAS) Cucumber alcohol
12	13.027	38331482	1.52	Pyrazine, methyl- (CAS) Methylpyrazine
13	13.292	49033980	1.95	1-Heptanamine, N-heptyl-N-nitro- (CAS) DIHEPTYL NITRAMINE
14	13.800	48746560	1.93	1,2-Hexadiene, 5-methyl- (CAS) 5-METHYL-1,2-HEXADIENE
15	14.294	100182931	3.97	1,2-DIMETHYLDIBORANE-D4
16	14.943	58918032	2.34	MONODEUTERO-1-HEXYNE
17	15.192	25553773	1.01	6-DIMETHYLAMINO-3,5-DIHYDROXY-1,2,4-TRIAZINE
18	15.57	1332448407	5.25	C11 ohexanone, 3-hydroxy- (CAS) 3-Hydroxycyclohexanone
19	16.219	55920181	2.22	-(6-OXA-BICYCLO[3.1.0]HEX-1-YL)-BUT-3-YN-2-ONE
20	16.544	95030755	3.77	Pinol, 2,6-dimethoxy- (CAS) 2,6-Dimethoxyphenol
21	16.742	30761807	1.22	2,2'-(1,4-PHENYLENE)BIS[4-(2,4-DICHLOROBENZYLIDENE)-4,5-DIHYDRO-5-
22	16.890	25346323	1.01	Pentane, 1-chloro- (CAS) 1-Chloropentane
23	16.992	23677774	0.94	Propanal, 2-methyl-, oxime
24	17.172	52992292	2.10	2-HYDROTY-1-ETHOXYETHYL-1-FURAN (NAME ?)
25	17.346	93298956	3.70	Pyrazine, 2-methoxy-6-methyl- (CAS) 2-Methoxy-6-methyl pyrazine
26	17.749	71507223	2.84	1 ETHYL-2-METHYL-PYRIDIN-4-YLAMINE
27	17.956	64859973	2.57	9-Oxabicyclo[6.1.0]non-4-ene (CAS) 1,2-Epoxy-5-cyclooctene
28	18.167	28806620	1.14	1-Butanamine, N-methyl-N-nitroso- (CAS) Butylmethylnitrosamine
29	18.371	86623565	3.44	FORMALDEHYDE, 1,1-DIDEUTERO-, 2,4-DINITROPHENYLHYDRAZON
30	18.77	1921523673	4.82	Butanal, 3-methyl- (CAS) 3-Methylbutanal
31	19.015	76398840	3.03	Piperazine, 2-methyl- (CAS) 2-Methylpiperazine
32	19.39	1431550732	5.22	Pregeijerene
33	20.05	1328072499	5.08	Silanediamine,1-chloro-N, N, N', N',1-pentamethyl-
34	20.390	83731457	3.32	16-Octadien-1-ol, 3,7-dimethyl- (CAS) 3,7-DIMETHYL 2,6-OCTADIENE-1-OL
35	20.651	46553815	1.85	3,3,3-TRIFLUORO-N-(2-FLUOROPHENYL)-2-
				(TRIFLUOROMETHYL)PROPIONAMIDE

Jurnal Aisyah: Jurnal Ilmu Kesehatan, 8 (3) 2023, - 1247

				5			
36	20.92	1458289271	6.28	6H-Pyrazolo[1,2-a][1,2,4,5]tetrazine, hexahydro-2,3-dimethyl- (CAS) 1,3,4,6-			
				TETRAAZABICYCLO			
37	21.497	21046659	0.83	Piperazine, 2-methyl- (CAS) 2-Methylpiperazine			
38	21.571	12075433	0.48	Bicyclo[2.2.1]heptan-2-one, 1,7,7-trimethyl-, oxime (CAS) 2-			
				HYDROXYIMINOBORNANE			
39	21.658	2606853	2.48	Cyclohexanone, 2-methyl-5-(1-methyl ethenyl)- (CAS) p-Menth-8-en-2-one			
40	22.181	61268768	2.43	1,4,5,8-TETRAAZADECALIN			
41	22542	11047993	0.44	7-Octynoic acid, methyl ester (CAS) METHYL-7-OCTYNOATE			
42	22.899	46778771	1.86	1-Piperazinecarboxylic acid, ethyl ester (CAS) Ethyl N-piperazinocarboxylate			
43	23.092	5968911	0.24	TRANS-8-EXOMETHOXYBICYCLO[4.3.0.]-3-NONENE-7-ENDO			
				CARBOXALDEHYDE			
44	23.167	15062454	0.60	Hepten-2-ol, (E)- (CAS) TRANS-HEPT-3-EN-2-OL			
45	23.392	6531738	0.26	2-(ALLYLAMINO)-5-ETHYL-1,3,4-THIADIAZOLE			
46	23.767	50112562	1.99	2,3-DIMETHYL-AZIRIDINE			
47	24.318	11365084	0.45	1-Aziridineethanol (CAS) 1-Azirdineethanol			
48	24.590	5973113	0.24	2,4-Pentanediamine, 2-methyl- (CAS) 2,4-DIAMINO-2-METHYLPENTANE			
49	26.860	4209413	0.17	1,3-Propanediamine, N, N-diethyl- (CAS) 3-DIETHYLAMINO PROPYL AMINE			
50	28.344	3639367	0.14	Phosphonic acid, diphenyl ester (CAS), Diphenyl phosphite			
51	29.271	66929662	2.66	3,7-Dimethyl-1,7-octadien-3-amine			
52	30.172	5573323	0.22	Ethyl N-methyl, N-ethylcarbamate			
53	30719	3606344	0.14	Benzenemethanol,.alpha(1-aminoethyl)-, (R*,R*)- (CAS) Norpseudoephedrine			
54	32.391	4329790	0.17	1,4,5,8-TETRAAZADECALIN			
55	35.189	3471638	0.14	1,3-Propanediamine, N, N-diethyl- (CAS) 3-DIETHYLAMINO PROPYL AMINE			
56	35.918	7256057	0.29	TETRACOSANE, 1-BROMO-			
57	36.267	8833182	0.35	N(2)-(t-Butoxycarbonyl)-N(6)-(benzyloxycarbonyl)-L-Lysin-4-acetamido anilide			
58	37.497	5077026	0.20	Pentanal (CAS) n-Pentanal			
59	38.142	15617318	0.62	Piperazine, 2-methyl- (CAS) 2-Methylpiperazine			
60	38.596	34894831	1.38	Vitamin E			
		2520711644	100				

10

The GC-MS analysis results of the extracts indicated variations in the percentage of each metabolite. The ethanol extracts of Pinang seeds contained thirty different compounds, while the fruits showed the presence of fifty compounds, and the leaves exhibited a total of sixty compounds. A noteworthy bioactive compound with antioxidant properties, Vitamin E, was detected at 0.20% in the fruits. In comparison, the analysis of Pinang leaves revealed a higher concentration of Vitamin E at 1.38%. Each of these bioactive compounds is associated with specific physiological functions. We conducted an α -glucosidase test using the Pinang seeds extract to explore the potential antidiabetic properties.

Alpha-Glucosidase Enzyme Inhibition

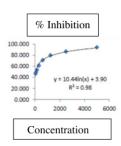
Numerous plant-based remedies have gained recognition for their potential in managing degenerative conditions such as diabetes, and indirectly, they serve as a source for developing prospective modern drugs, particularly for addressing hyperlipidemia. Bioactive compounds present in these plants often act as antioxidants, capable of impeding glucose absorptib. In vitro analyses have confirmed the α -glucosidase inhibitory properties of ethanolic-maceration extracts. These liver glucosidase inhibitors target -1, 6-glucosidase, a glycogen-debrancing enzyme within the liver. By doing so, they reduce the glycogenolytic rate, leading to increased glycogen storage tips enzyme within the liver. hibiting these enzyme systems results in decreased blood glucose levels, representing a short-term effect and a modest reduction in hemoglobin A1c levels. Given the various side effects associated with synthetic glucosidase inhibitors, researchers in the present era have a growing focus on herbal medicines (Papuc et al., 2017).

Therefore, the Pinang seeds extracts (in 80% ethanol) showed a potent α -glycosidase inhibitor in this study. The α -glucosidase inhibition of seeds may be 2 ated to the bioactive compounds properties of the fruits and leaves of this plant (Yin *et al.*, 2014). The benefit of these inhibitors would retard starch digestion, absorption, and lowering blood glucose levels. IC₅₀ values of α -glucosidase inhibited seeds extract were 82.74 ppm, and the global standard was 0.34 ppm, presented in Figures 5 and 6. This research showed an inhibitory of the α -glucosidase activity of seeds of Pinang. Our results suggest that the extract of Pinang seeds is a potential candidate for developing antidiabetic agents.

Potential Antimicrobial Effect

The data indicates that the extract derived from Pinang seeds has demonstrated potential antimicrobial properties by inhibiting the growth of the Gram-negative bacterium Escherichia coli ATCC 25922, as presented in Table 4. Our findings suggest that the mechanism of action of the Pinang seeds extract against E. coli could involve membrane depolarization and increased membrane permeability, which affect intracellular enzyme activities and elevate intracellular levels of reactive oxygen species (ROS). These alterations may lead to cell apoptosis and, ultimately, the death of the bacteria (Shah et al., 2018; Zhang et al., 2020; Lin et al., 2019).

Jurnal Aisyah: Jurnal Ilmu Kesehatan, 8(3) 2023, - 1248



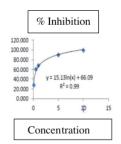


Figure 5. Values Inhibition of Pinang seeds

Figure 6. Values Inhibition of Acarbose

Table 4
Potential antimicrobial effect

Sample	Condition	Parameter		Result	Unit	Analysis
		E. Coli	10%	7.38	mm	diffusion
		antibacterial	5%	6.00	mm	diffusion
Pinang seeds	solid		2.5%	6.00	mm	diffusion
		S. Aureus	10%	6.00	mm	diffusion
		antibacterial	5%	6.00	mm	diffusion
			2.5%	6.00	mm	diffusion
Control (+)		E. Coli	0.1%	22.5	mm	diffusion
Chloram	solid	antibacterial				
Phenicol		S. Aureus antibacterial	0.1%	24.14	mm	diffusion
Control (-)		E. Coli	20%	6.00	mm	diffusion
DMSO	liquid	antibacterial				
	•	S. Aeureus antibacterial	20%	6.00	mm	diffusion

Disc diameter: 6.00 mm

The results of the GC-MS analysis of the seeds, fruits, and leaves of Pinang (*Areca catechu* L.) plants are essential to determine the organic compounds. Vitamin E in fruits and leaves acts as an antioxidant in this plant. Pinang seeds' concentration of bioactive compounds has potential α-glucosidase inhibited activity, impacting the delayed absorption of carbohydrates and the potent effect on diabetes treatment to increase health 11 ucose blood levels. The Pinang seeds formula has the potential antimicrobial activity to inhibit the growth of *E.coli* bacteria as it showed zone inhibition. The bioactive compounds in this plant extract might prevent diabetes and antimicrobial and may serve as alternative drugs for treating various illnesses in human beings.

CONCLUSIONS AND SUGGESTIONS

The analysis results of bioactive components in seeds, fruits, and leaves 10 Areca catechu (Areca catechu L.) plants have antioxidant activity. Active components in Areca nut seeds have the ability to inhibit the activity of the enzyme alpha-glucosidase, which affects carbohydrate metabolism and has the potential for alternative treatment of chronic hyperglycemia. Areca nut seeds have antibacterial bioactivity against Gram-negative bacteria E.coli, with an inhibition zone. Further research is needed to determine the benefits of the chemical composition of the Areca nut plant as an alternative medicine to help maintain health.

2 ACKNOWLEDGEMENT

The authors thank the Faculty of Medicine, Universitas Kristen Indonesia, for funding this project. Our special thanks to all staff members in the Laboratory of Biochemistry, Laboratory of Microbiology, Laboratory of Parasitology Medical Faculty of UKI Jakarta, and Laboratory of Physiology IPB Bogor.

REFERENCES

- Alqahtani, A. S., Hidayathulla, S., Rehman, M. T., ElGamal, A. A., Al-Massarani, S., Razmovski-Naumovski, V., ... & AlAjmi, M. F. (2019). Alpha-amylase and alpha-glucosidase enzyme inhibition and antioxidant potential of 3-oxolupenal and katononic acid isolated from Nuxia oppositifolia. *Biomolecules*, 10(1), 61.
- Ansari, P., Akther, S., Hannan, J. M. A., Seidel, V., Nujat, N. J., & Abdel-Wahab, Y. H. (2022). Pharmacologically active phytomolecules isolated from traditional antidiabetic plants and their therapeutic role for the management of diabetes mellitus. *Molecules*, 27(13), 4278.
- Arya, A., Al-Obaidi, M. M. J., Karim, R. B., Taha, H., Khan, A. K., Shahid, N., ... & Ali, H. M. (2015). Extract of Woodfordia fruticosa flowers ameliorates hyperglycemia, oxidative stress and improves β-cell function in streptozotocin–nicotinamide induced diabetic rats. *Journal of ethnopharmacology*, 175, 229-240.
- Aschner, P., Karuranga, S., James, S., Simmons, D., Basit, A., Shaw, J. E., ... & Saeedi, P. (2021). The International Diabetes Federation's guide for diabetes epidemiological studies. *Diabetes research and clinical practice*, 172.
- Chen, H., & Li, R. (2021). Introduction of Diabetes Mellitus and Future Prospects of Natural Products on Diabetes Mellitus. Structure and Health Effects of Natural Products on Diabetes Mellitus, 1-15.
- Chipiti, T., Ibrahim, M. A., Singh, M., & Islam, M. S. (2015). In vitro α-amylase and α-glucosidase inhibitory effects and cytotoxic activity of Albizia antunesiana extracts. *Pharmacognosy Magazine*, 11(Suppl 2), S231.
- Dan, W. J., Zhang, Q., Zhang, F., Wang, W. W., & Gao, J. M. (2019). Benzonate derivatives of acetophenone as potent α-glucosidase inhibitors: Synthesis, structure–activity relationship and mechanism. *Journal of Enzyme Inhibition* and Medicinal Chemistry, 34(1), 937-945.
- Demir, S., Nawroth, P. P., Herzig, S., & Ekim Üstünel, B. (2021). Emerging targets in type 2 diabetes and diabetic complications. *Advanced Science*, 8(18), 2100275.
- Di Santo, M. C., D'Antoni, C. L., Rubio, A. P. D., Alaimo, A., & Pérez, O. E. (2021). Chitosan-tripolyphosphate nanoparticles designed to encapsulate polyphenolic compounds for biomedical and pharmaceutical applications— A review. *Biomedicine & Pharmacotherapy*, 142, 111970.
- Doan, H. V., Riyajan, S., Iyara, R., & Chudapongse, N. (2018). Antidiabetic activity, glucose uptake stimulation and α-glucosidase inhibitory effect of Chrysophyllum cainito L. stem bark extract. *BMC Complementary and Alternative Medicine*, 18(1), 1-10.
- Doan, H. V., Riyajan, S., Iyara, R., & Chudapongse, N. (2018). Antidiabetic activity, glucose uptake stimulation and α-glucosidase inhibitory effect of Chrysophyllum cainito L. stem bark extract. *BMC Complementary and Alternative Medicine*, 18(1), 1-10.
- Ha, K. S., Jo, S. H., Mannam, V., Kwon, Y. I., & Apostolidis, E. (2016). Stimulation of phenolics, antioxidant and α-glucosidase inhibitory activities during barley (Hordeum vulgare L.) seed germination. *Plant Foods for Human Nutrition*, 71, 211-217.
- Hu, W., Li, C., Dai, J., Cui, H., & Lin, L. (2019). Antibacterial activity and mechanism of Litsea cubeba essential oil against methicillin-resistant Staphylococcus aureus (MRSA). *Industrial Crops and Products*, 130, 34-41.
- Hussain, S., & Chowdhury, T. A. (2019). The impact of comorbidities on the pharmacological management of type 2 diabetes mellitus. *Drugs*, 79(3), 231-242.
- Iwu, I. C., Chijioke-okere, M., Onu, U. L., & Uchegbu, R. (2018). GC-MS, Phytochemical and Antimicrobial Analysis of the Leaf of Newboudia laevis P. Benth. *International Journal of Innovative Research and Development*. 2018a, 7(7), 242-250.
- Jeong, D., & Priefer, R. (2022). Anti-obesity weight loss medications: Short-term and long-term use. Life Sciences, 120825.
- Kachangoon, R., Vichapong, J., Santaladchaiyakit, Y., & Srijaranai, S. (2020). Cloud-point extraction coupled to in-situ metathesis reaction of deep eutectic solvents for preconcentration and liquid chromatographic analysis of neonicotinoid insecticide residues in water, soil and urine samples. *Microchemical Journal*, 152, 104377.
- Lin, Y., Tang, X., Xu, L., & Wang, S. (2019). Antibacterial properties and possible action mechanism of chelating peptides-zinc nanocomposite against Escherichia coli. Food Control, 106, 106675.
- Nabi, M., Tabassum, N., & Ganai, B. A. (2022). Skimmia anquetilia NP Taylor and airy shaw (rutaceae): A critical appriasal of its ethnobotanical and pharmacological activities. *Frontiers in Plant Science*, 13, 930687.
- Papuc, C., Goran, G. V., Predescu, C. N., Nicorescu, V., & Stefan, G. (2017). Plant polyphenols as antioxidant and antibacterial agents for shelf-life extension of meat and meat products: Classification, structures, sources, and action mechanisms. Comprehensive Reviews in Food Science and Food Safety, 16(6), 1243-1268.
- Ren, X., An, P., Zhai, X., Wang, S., & Kong, Q. (2019). The antibacterial mechanism of pterostilbene derived from xinjiang wine grape: A novel apoptosis inducer in Staphyloccocus aureus and Escherichia coli. Lwt, 101, 100-106.
- Saad, B., Kmail, A., & Haq, S. Z. (2022). Anti-diabesity Middle Eastern medicinal plants and their action mechanisms. Evidence-Based Complementary and Alternative Medicine, 2022.

Jurnal Aisyah: Jurnal Ilmu Kesehatan, 8(3) 2023, - 1250

- Schauer, P. R., Bhatt, D. L., Kirwan, J. P., Wolski, K., Aminian, A., Brethauer, S. A., ... & Kashyap, S. R. (2017). Bariatric surgery versus intensive medical therapy for diabetes—5-year outcomes. *New England Journal of Medicine*, 376(7), 641-651.
- Shah, S. R., Ukaegbu, C. I., Hamid, H. A., & Alara, O. R. (2018). Evaluation of antioxidant and antibacterial activities of the stems of Flammulina velutipes and Hypsizygus tessellatus (white and brown var.) extracted with different solvents. *Journal of Food Measurement and Characterization*, 12, 1947-1961.
- Shahvar, A., Shamsaei, D., & Saraji, M. (2020). A portable smartphone-based colorimetric sensor for rapid determination of water content in ethanol. *Measurement*, 150, 107068.
- Sharma, A. K., Kumar, A., Yadav, S. K., & Rahal, A. (2014). Studies on antimicrobial and immunomodulatory effects of hot aqueous extract of Acacia nilotica L. leaves against common veterinary pathogens. *Veterinary medicine* international, 2014.
- Yin, Z., Zhang, W., Feng, F., Zhang, Y., & Kang, W. (2014). α-Glucosidase inhibitors isolated from medicinal plants. Food Science and Human Wellness, 3(3-4), 136-174.
- Zhang, L. L., Zhang, L. F., & Xu, J. G. (2020). Chemical composition, antibacterial activity and action mechanism of different extracts from hawthorn (Crataegus pinnatifida Bge.). Scientific reports, 10(1), 8876.
- Zhang, L. L., Zhang, L. F., & Xu, J. G. (2020). Chemical composition, antibacterial activity and action mechanism of different extracts from hawthorn (Crataegus pinnatifida Bge.). *Scientific reports*, 10(1), 8876.

Turn it in Bio active compounds antidia beticand antimic robial p...

ORIGIN	ALITY REPORT		<u>'</u>
SIMIL	4% 11% ARITY INDEX INTERNET SOURCES	9% PUBLICATIONS	4% STUDENT PAPERS
PRIMAF	Y SOURCES		
1	www.knowitall.com Internet Source		2%
2	repository.uki.ac.id Internet Source		2%
3	premierpublishers.org Internet Source		1 %
4	thescipub.com Internet Source		1 %
5	psg.gsfc.nasa.gov Internet Source		1 %
6	www.ijddr.in Internet Source		1 %
7	www.phytopharmajou	rnal.com	1 %
8	bmccomplementmedth Internet Source	nerapies.biome	dcentral.com 1 %
9	eprints.unm.ac.id Internet Source		1%

Exclude quotes On Exclude matches < 1%

Exclude bibliography On